

CENTER FOR NEUROSCIENCE AND CELL BIOLOGY UNIVERSITY OF COIMBRA PORTUGAL

Exploring high-content screening as a functional genomics tool in biomedicine

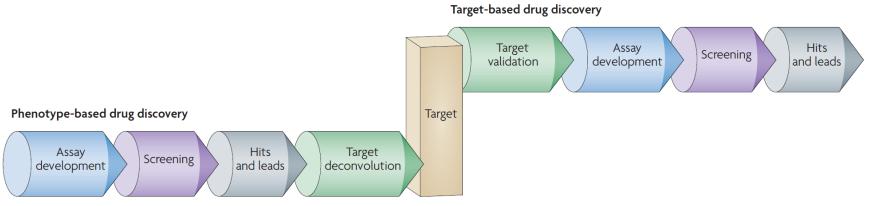
Miguel Mano

Functional Genomics and RNA-based Therapeutics Laboratory

ICGEB-TRAIN Workshop, Bled, Slovenia | May 2019

Target identification	Target assay construction and screening	Target validation, hits-to-leads medicinal chemistry safety	Phase I: safety	Phase II: efficacy, safety	Phase III: efficacy, safety	FDA review and approval
					1	

Roses, Nature Reviews Drug Discovery 2008



Terstappen et al., Nature Reviews Drug Discovery 2007

Functional genomics

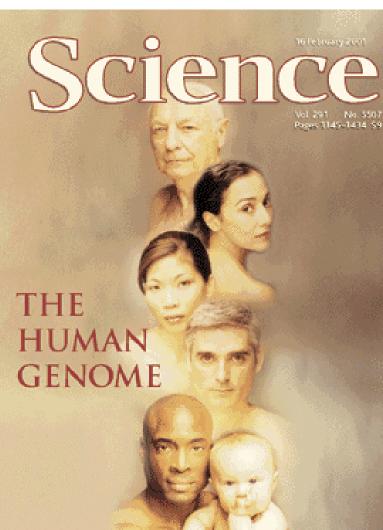
Application of global (genome-wide or system-wide) experimental approaches to assess gene function

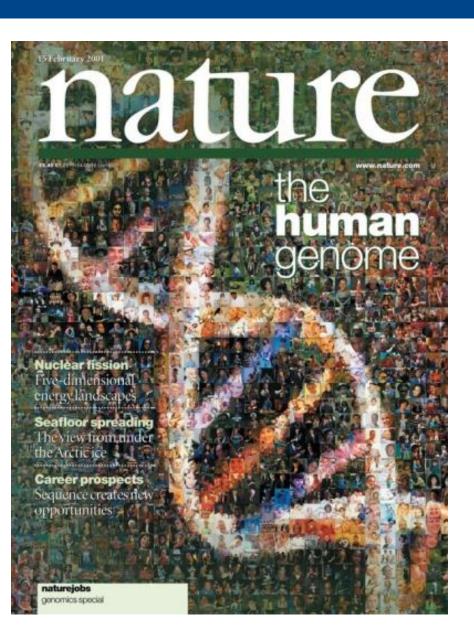
Genome Sequencing



S. cerevisiae 1997 C. elegans 1998 D. melanogaster 2000 A. thaliana 2000 H. sapiens 2001 M. musculus 2002 R. norvegicus 2004 (...)

The Human Genome at 18 | Feb.2001-Feb.2019







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STATISTICS.

No. of Concession, name

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haploid genome, font size 5

The Human Genome in numbers

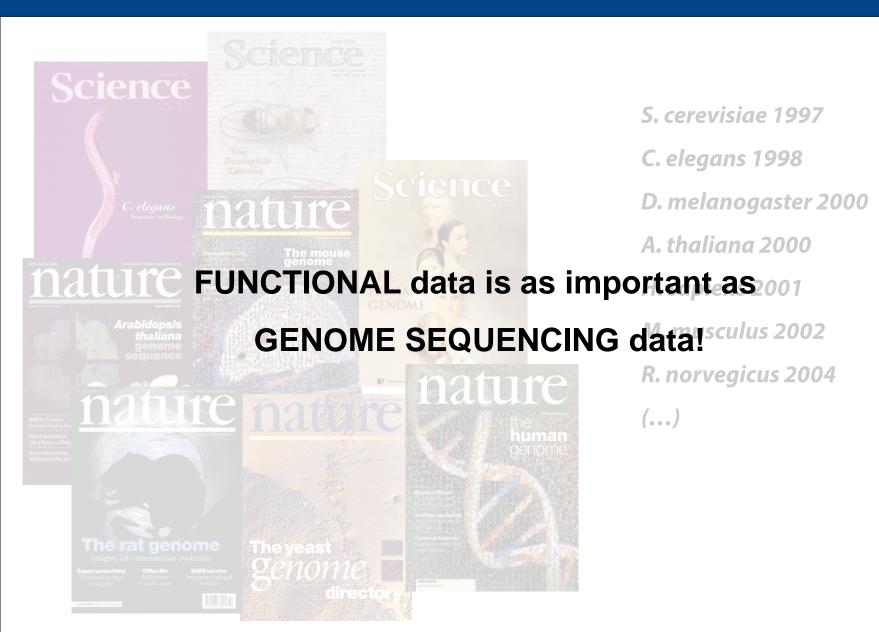
23 chromosome pairs

~ 6 500 000 000 base pairs 2-3 meters in lenght

~ 20 000 protein coding genes

< 2% genome encodes proteins

Genome Sequencing



Functional genomics

Application of global (genome-wide or system-wide) experimental approaches to assess gene function

How can we study gene function?

Expression analysis

- Analysis of mRNAs/microRNAs/IncRNAs present at different stages
- Characterization changes occurring after different stimuli

Functional screening

- Analysis of the functional impact of the perturbation of a single protein/microRNA/lncRNA
- Relevance of proteins/microRNAs/IncRNAs to a given function/ /cellular process/phenotype

Expression analysis

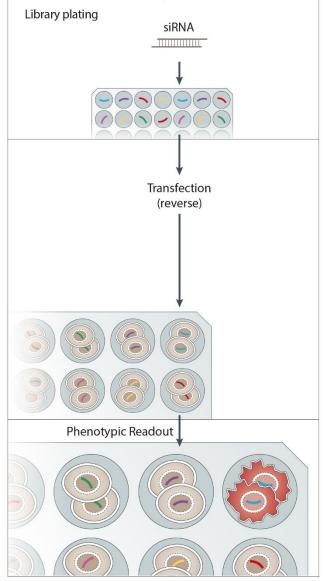
- DNA/RNA sequencing (mRNA, ncRNAs)
- **Proteomics** (proteins)
- Lipidomics (lipids)
- ...omics

Functional screening

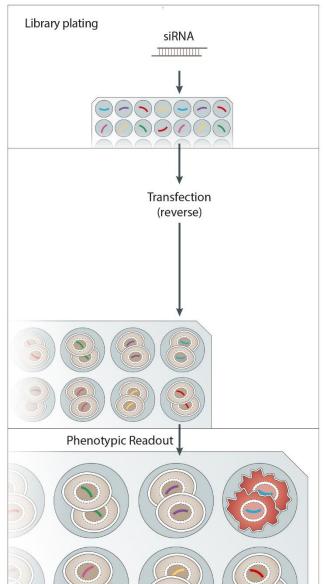
- Gene silencing (RNAi)
- Gene overexpression (expression libraries)
- Gene knockout (CRISPR)
- miRNA modulation

Phenotypic Screening in Functional Genomics

ARRAYED SCREENINGS



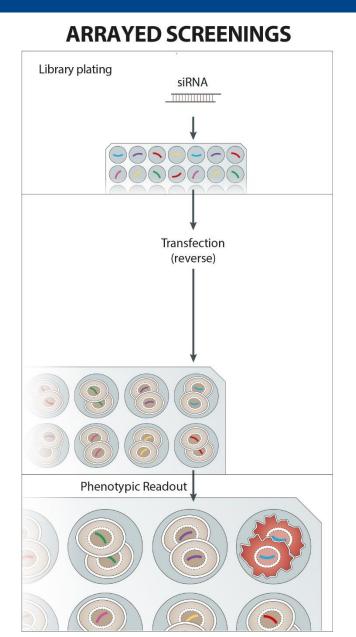
Phenotypic Screening in Functional Genomics



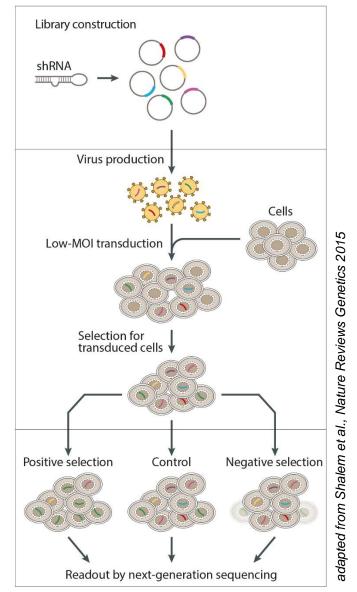
ARRAYED SCREENINGS

siRNAs microRNA mimics microRNA inhibitors

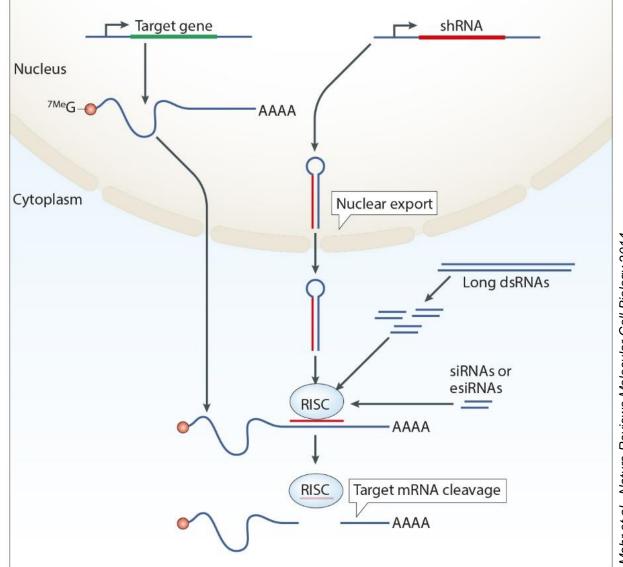
Phenotypic Screening in Functional Genomics



POOLED SCREENINGS



RNA interference



Mohr et al., Nature Reviews Molecular Cell Biology 2014

Homogeneous assays vs. Microscopy-based readouts

original image

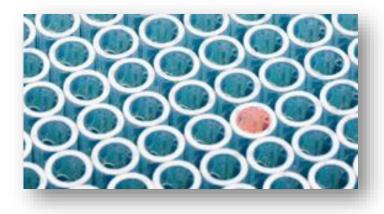


Plate-reader Readouts High-Throughput Screening

Simple optimization Fast Quantitative

PROS

Microscopy Readouts High-Content Screening Ouantitative

analysis

Quantitative Multiparametric Rich biological information Subcellular resolution

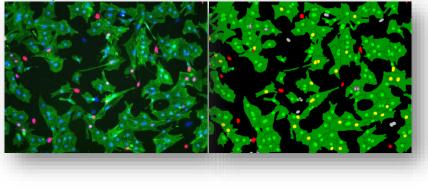
Binary results Poor biological information

CONS

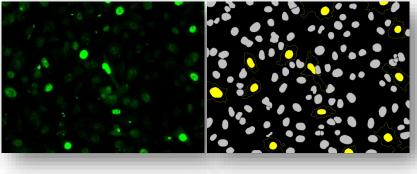
Slow (acquisition and analysis) Complex optimization Complex analysis

High-Content Screening: getting the whole picture

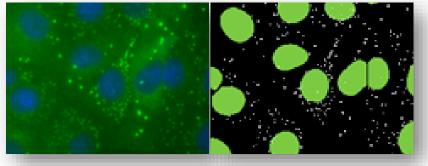
Cell proliferation



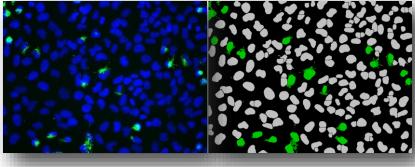
Nucleo-cytoplasmic translocation

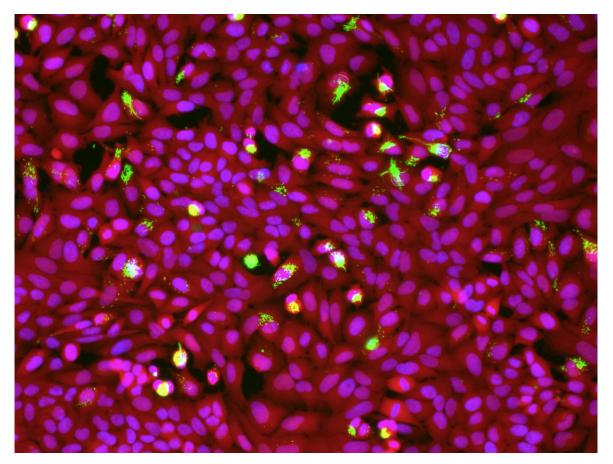


Viral infection

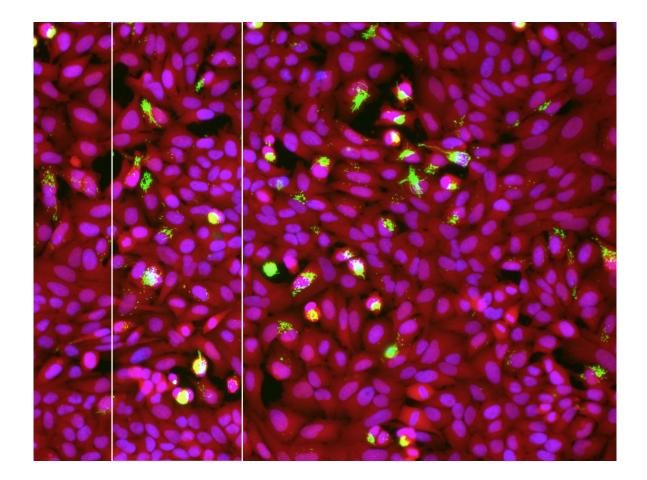


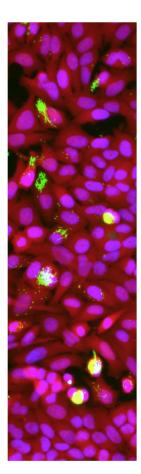
Bacterial infection

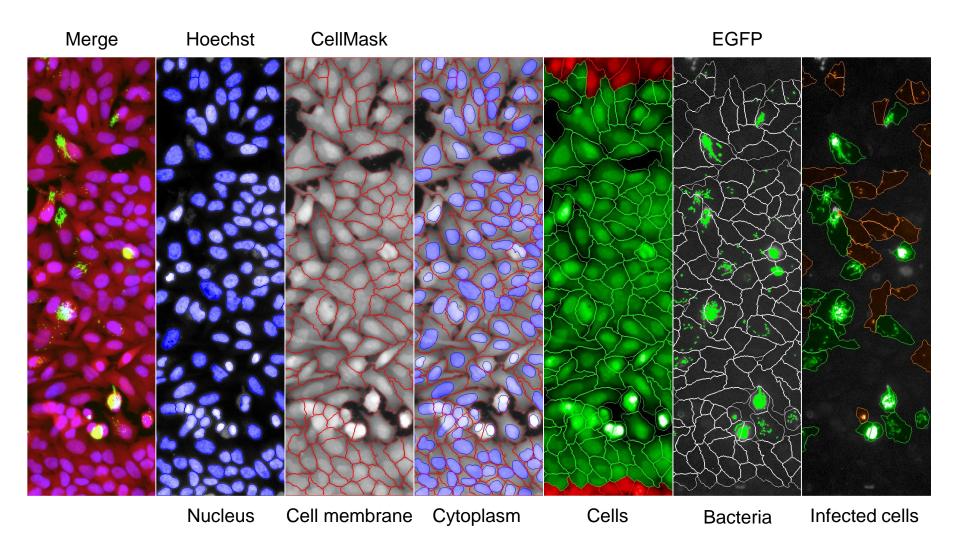


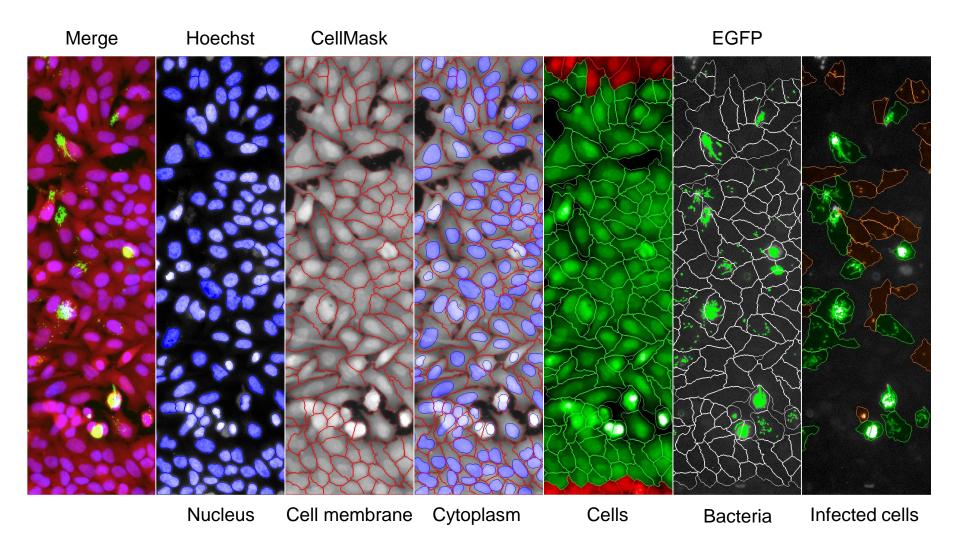


Hoechst Salmonella Typhimurium CellMask









Unbiased extraction of multiparametric numerical data from images



Functional Genomics and RNA-based Therapeutics laboratory

CNC/UC-Biotech | University of Coimbra, Portugal

 Fully automated state-of-the-art high-throughput/high-content screening platform for functional screenings in human and mouse cellular models (BSL2 environment)

• Libraries:

human genome-wide siRNA (18,175 gene targets; 4 siRNAs + pools) mouse genome-wide siRNA (16,872 gene targets; 4 siRNAs + pools) human microRNA mimics (miRBase v21; 2,588 mature sequences) human microRNA inhibitors (miRBase v21; 2,588 mature sequences) FDA approved drugs (640 chemical compounds)

• Technologies:

High-throughput screening High-content microscopy screening Live-cell microscopy screening



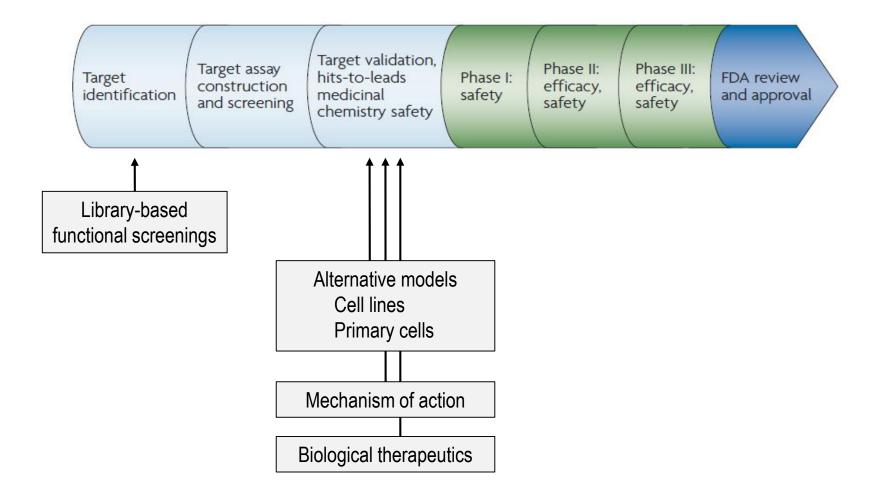
Miniaturization: costs and benefits

	6-well	96-well	384-well	1536-well	
			-	1.5mm •	
Well area (cm ²)	9.5	0.3	0.06	0.02	
Number cells	1x10 ⁶	3x10 ⁴	8x10 ³	2x10 ³	
Working volume	2-3ml	100-200µl	25-50µl	5-10µl	

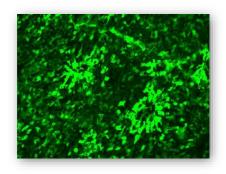
Assay complexity

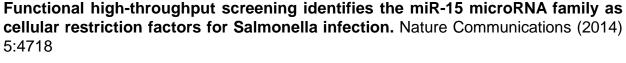
Throughput

Functional genomics screening as a tool for the development of novel therapeutics



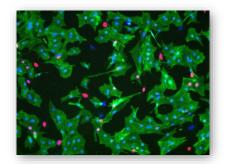
Functional screening: examples of applications





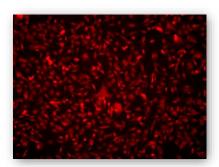
Genome-wide RNAi screening identifies host restriction factors critical for in vivo AAV transduction. PNAS (2015) 112:11276

Analysis of host microRNA function in Shigella infection reveals a role for miR-29b-2-5p in bacterial capture by filopodia. PLOS Pathogens (2017) 13:e1006327 Cellular TRIM33 restrains HIV-1 infection by targeting viral integrase for proteasomal degradation. Nature Communications (2019) 10:926



Functional screening identifies miRNAs inducing cardiac regeneration. Nature (2012) 492:376

Id genes are essential for early heart formation. Genes and Development (2017) 31:1325



Metabolic control of YAP and TAZ by the mevalonate pathway. Nature Cell Biology (2014) 16:357

Glucocorticoid receptor signalling activates YAP in breast cancer. Nature Communications (2017) 8:14073

High-throughput screening uncovers miRNAs enhancing glioblastoma cell susceptibility to tyrosine kinase inhibitors. Human Molecular Genetics (2017) 26:4375-4387

Mechanical cues control mutant p53 stability through a Mevalonate/RhoA axis. Nature Cell Biology (2018) 20:28-35

Sterol Regulatory Element Binding Protein 1 couples mechanical cues and lipid metabolism. Nature Communications (2019) 10:1326

High-Content Screening

How high is high?

Homogeneous assays vs. Microscopy-based readouts

original image

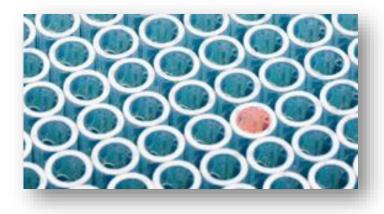


Plate-reader Readouts High-Throughput Screening

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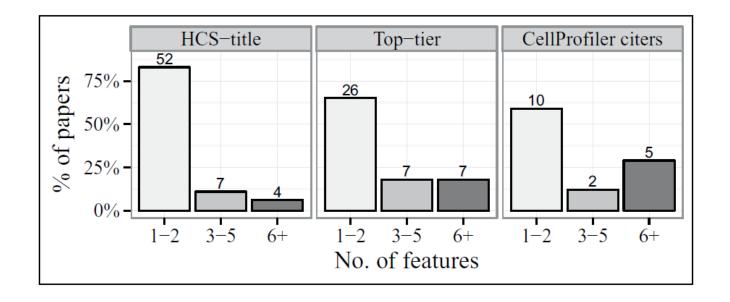
Quantitative Multiparametric Rich biological information Subcellular resolution

Binary results Poor biological information

CONS

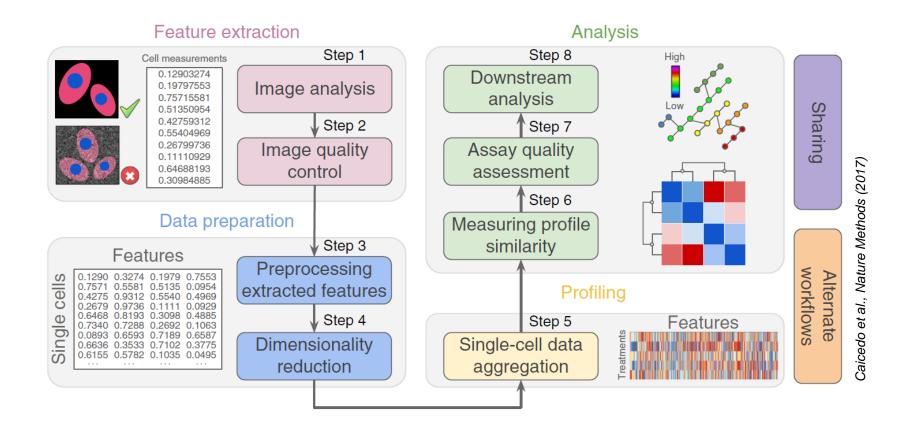
Slow (acquisition and analysis) Complex optimization Complex analysis

High-content screening... really?



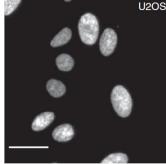
"... although the number of HCS experiments published each year continues to grow steadily, the information content lags behind. We find that a majority of high-content screens published so far (60–80%) made use of only one or two image-based features measured from each sample and disregarded the distribution of those features among each cell population."

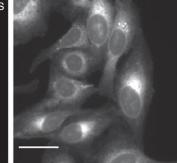
Image-based cell profiling

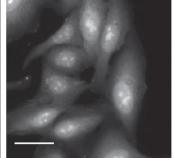


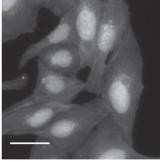
- Global profiling of (genetic, chemical) perturbations based on multiparametric analysis of cell morphology
- Typically involves multiplexed labeling (fluorescent dyes, antibodies) of subcellular structures/organelles

Cell Painting

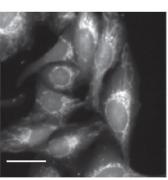








Actin, Golgi, plasma membrane



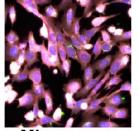
Nucleus

Endoplasmic reticulum

Nucleoli, cytoplasmic RNA

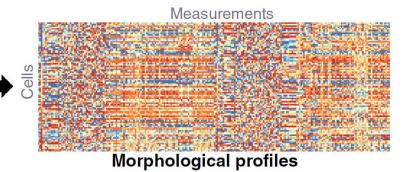
Mitochondria

Bray et al., Nature Protocols (2016)



Microscopy imaging

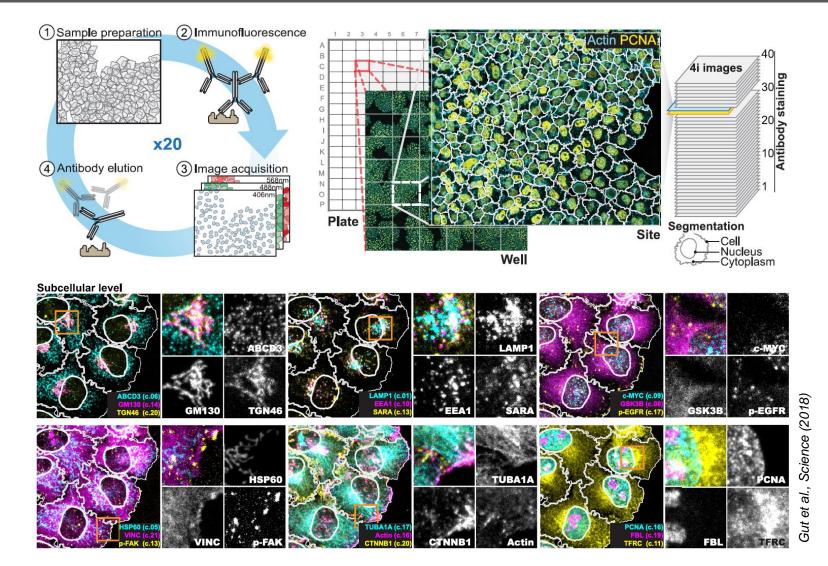
Image analysis



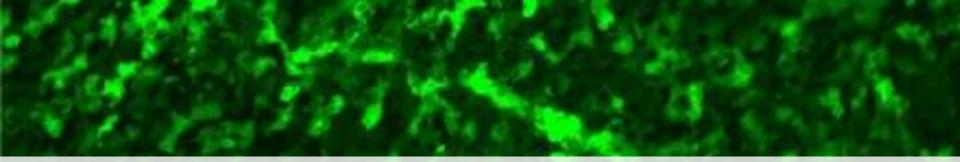
Caicedo et al., Current Opinion in Biotechnology (2016)

- 6 fluorescent dyes (no antibodies)
- 8 subcellular structures/organelles
- 5 channels
- ~1,500 morphological measurements (features) per cell

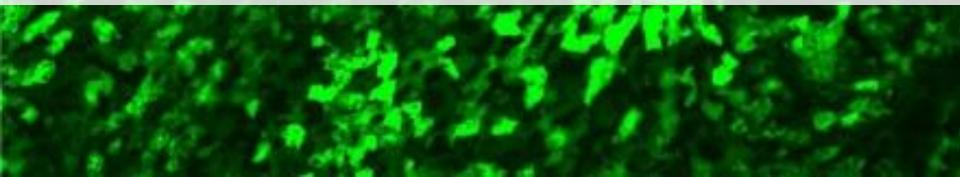
High-throughput multiplexed protein readouts iteractive indirect immunofluorescence imaging



40 off-the-shelf antibodies (20 cycles); ~4h per cycle

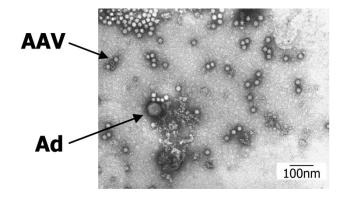


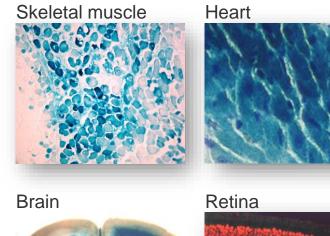
GENOME-WIDE SIRNA SCREENING Identification of host cell factors controlling transduction of vectors based on Adeno-Associated Virus (AAV)

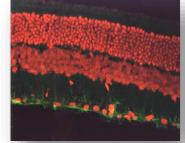


Adeno-Associated Virus (AAV) vectors

- Vectors based on AAV are efficient gene transfer tools
- AAV vectors transduce differentiated, post-mitotic tissues with high efficiency
- AAV vectors permit a sustained, long term persistence of the transgene in vivo without integrating into the host genome
- AAV vectors can be prepared at high titers and purity
- Expression of the gene of interest can be driven by any desirable promoter
- Outcome of transduction is critically dependent on the interplay between vectors and the host cell







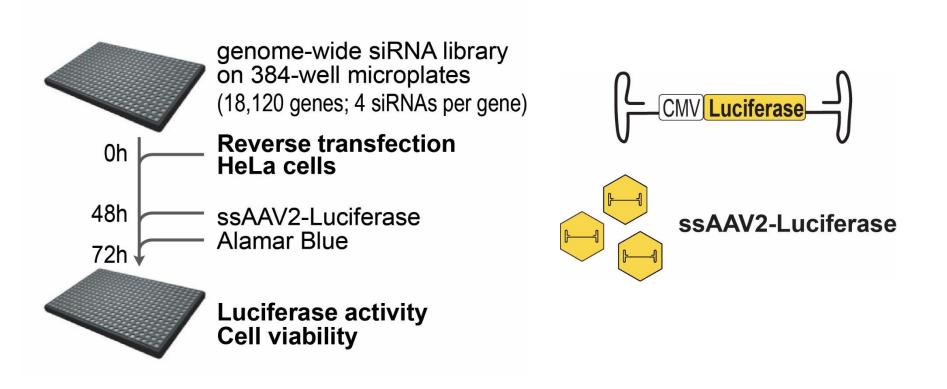
Genome-wide siRNA screening for AAV transduction

Main Goals

- Identify host cell factors controlling transduction by AAV vectors
- Improve current knowledge of the intracellular processing of AAV vectors
- Improve AAV as vectors for gene therapy
 Broaden the cell/tissue tropism of AAV vectors
 Increase efficiency of AAV vectors
 Decrease AAV vector dosage

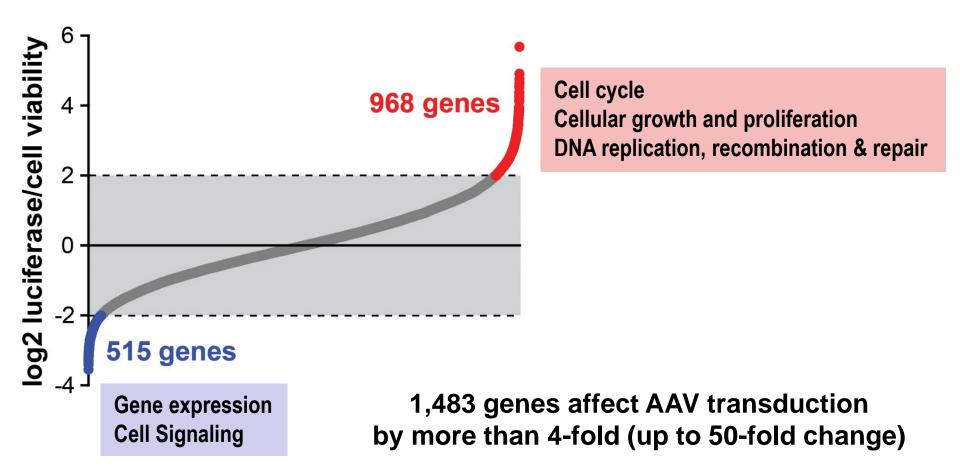
Genome-wide siRNA screening for AAV transduction

Experimental Workflow



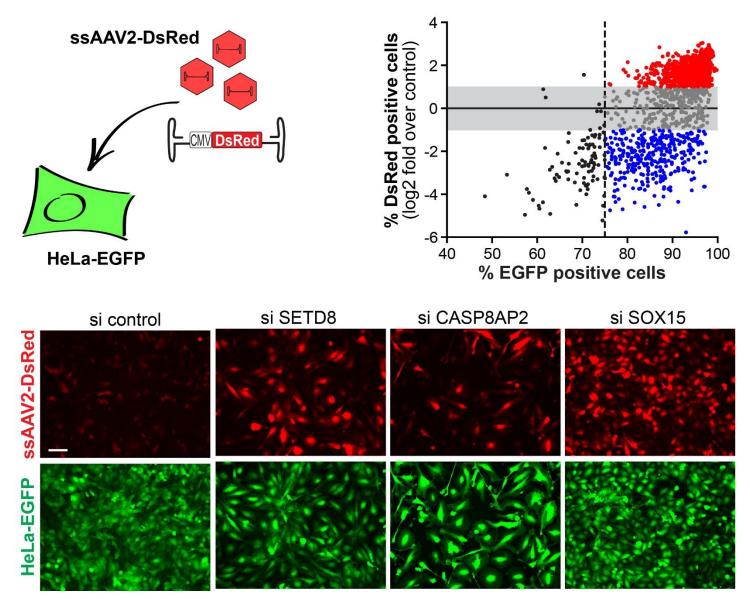
Genome-wide siRNA screening for AAV transduction

18,120 GENE TARGETS



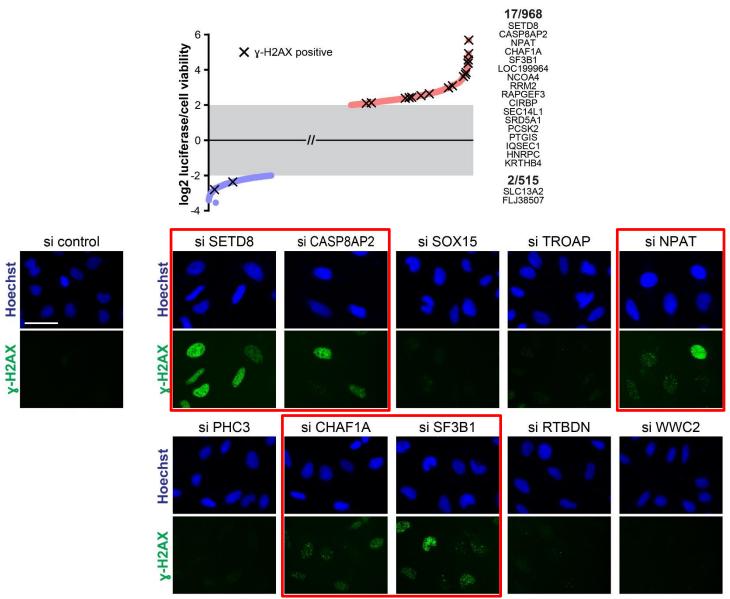
178 genes by more than 8-fold

Secondary screening – Refinement for AAV-specific targets



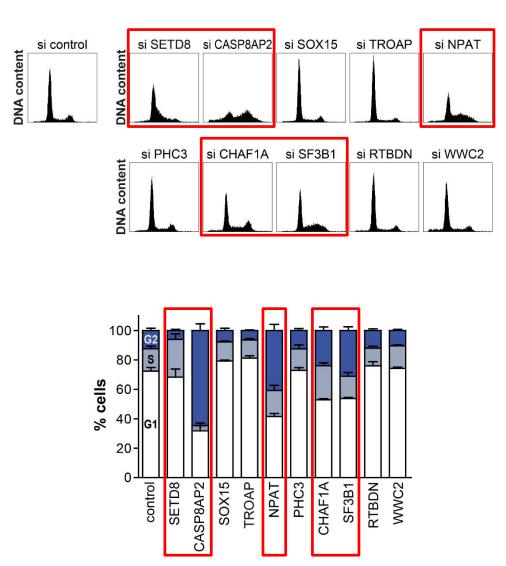
4 fields per well, 10x magnification, average 1,500 cells analyzed per well

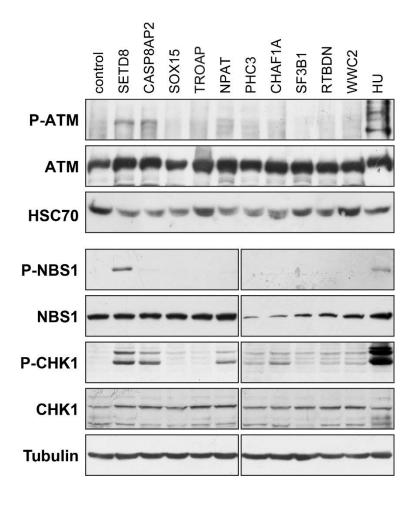
Secondary screening – vH2AX foci



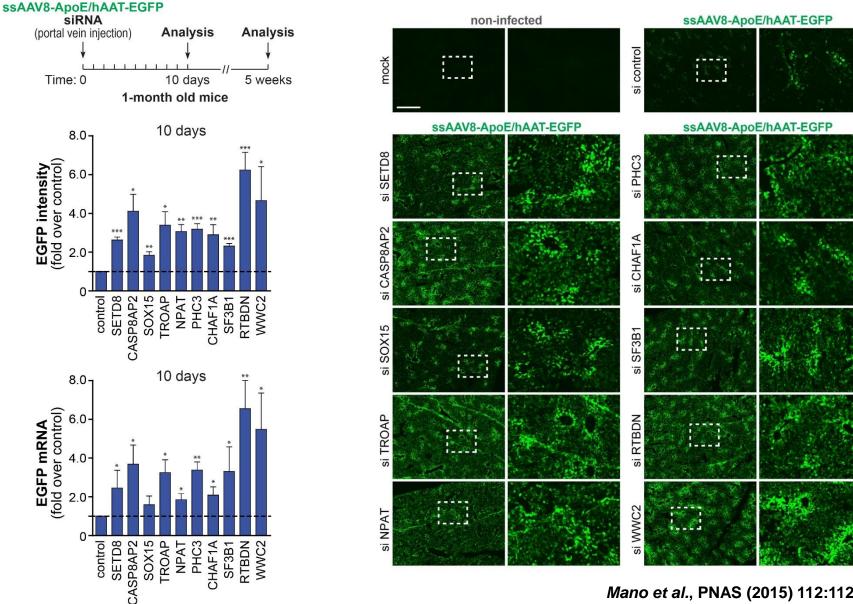
9 fields per well, 20x magnification

DNA Damage Response – Top 10 gene targets

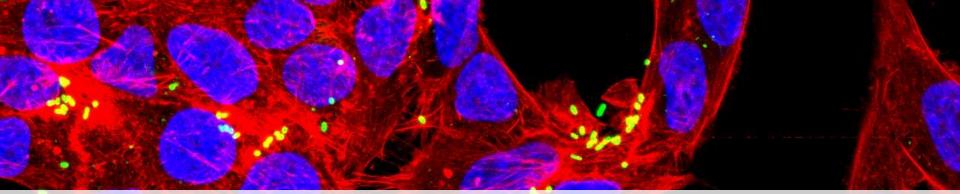




Effect of siRNAs on AAV transduction in vivo (liver)



Mano et al., PNAS (2015) 112:11276



microRNA SCREENING

microRNAs controlling infection by bacterial pathogens

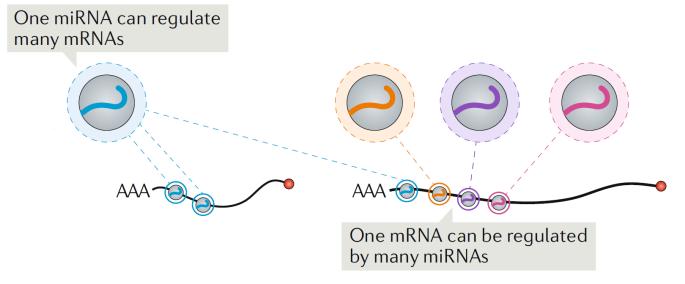
in collaboration with Ana Eulalio lab

Institute for Molecular Infection Biology | University of Würzburg, Germany Center for Neuroscience and Cell Biology | University of Coimbra, Portugal

MicroRNAs

MicroRNAs are genome-encoded (20-22nt) non-coding RNAs that play a major role in the post-transcriptional regulation of gene expression

More than 2,500 human mature miRNAs currently annotated

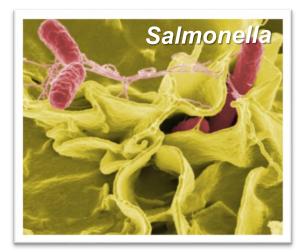


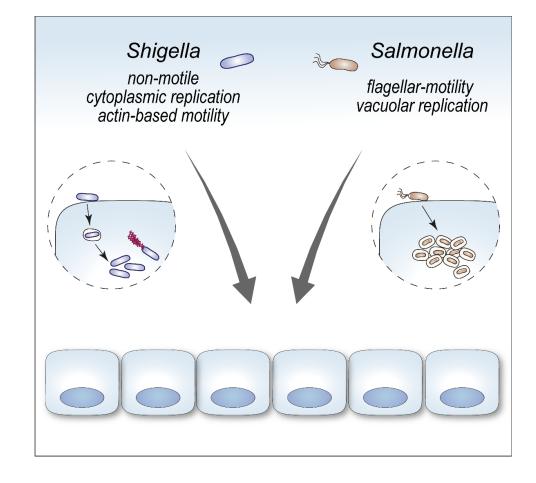
adapted from Gebert & MacRae. 2018. Nat Rev Mol Cell Biol.

MicroRNAs have been shown to be involved in a broad range of biological processes

Shigella flexneri and Salmonella Typhimurium

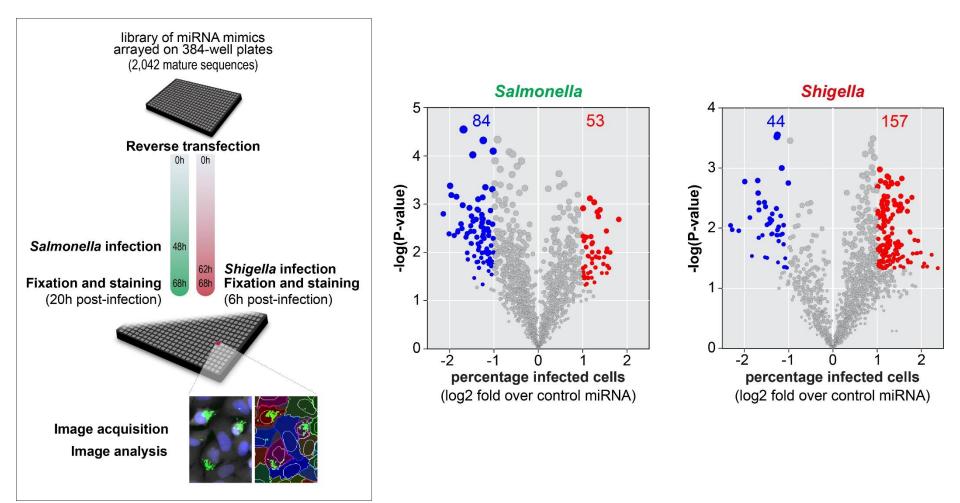




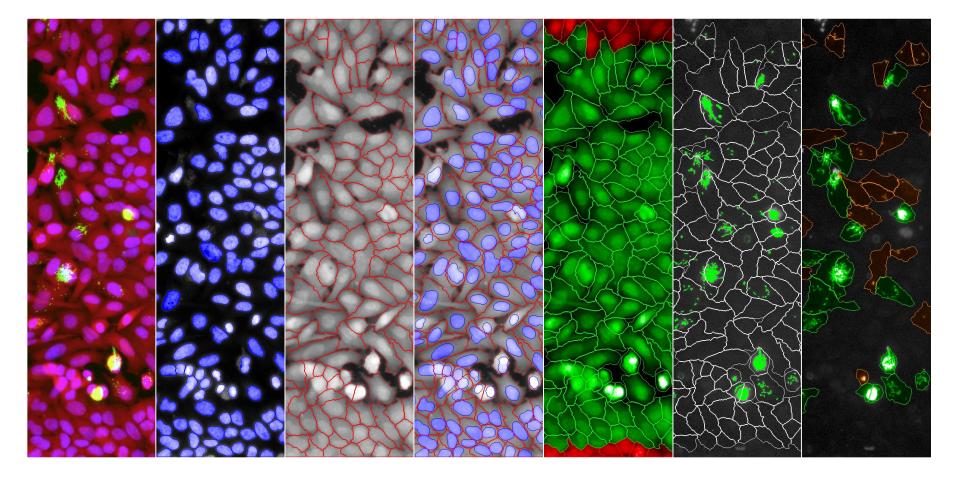


Among the most important bacterial pathogens causing food-borne diseases

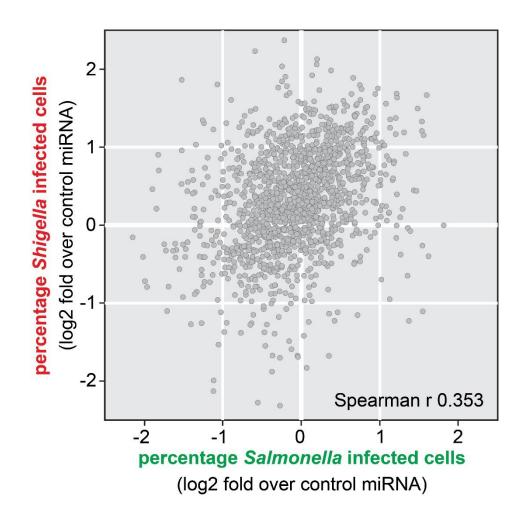
Host miRNAs are major regulators of Salmonella and Shigella infection

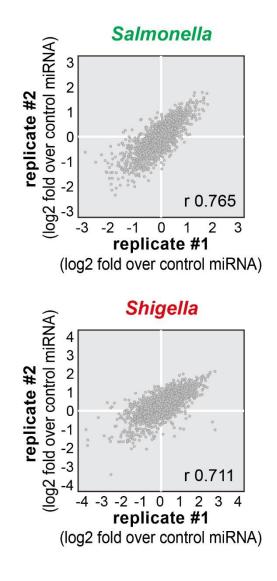


Unbiased extraction of multiparametric numerical data from images Subcellular resolution, single-cell analysis



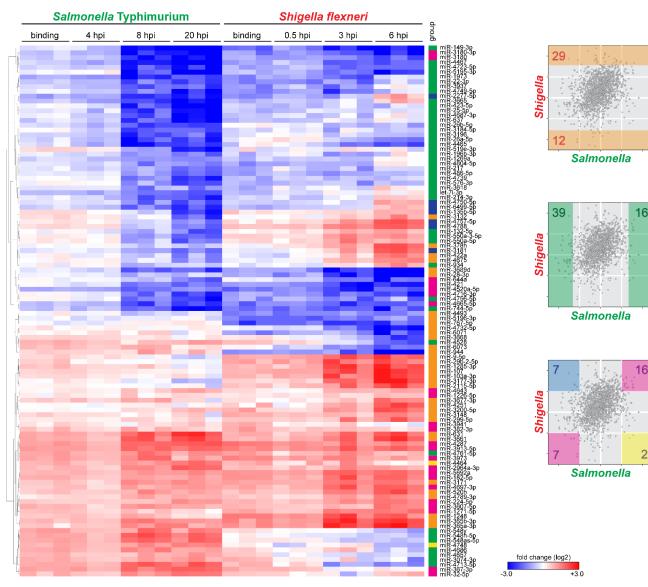
Salmonella and Shigella infection are regulated by a different set of miRNAs





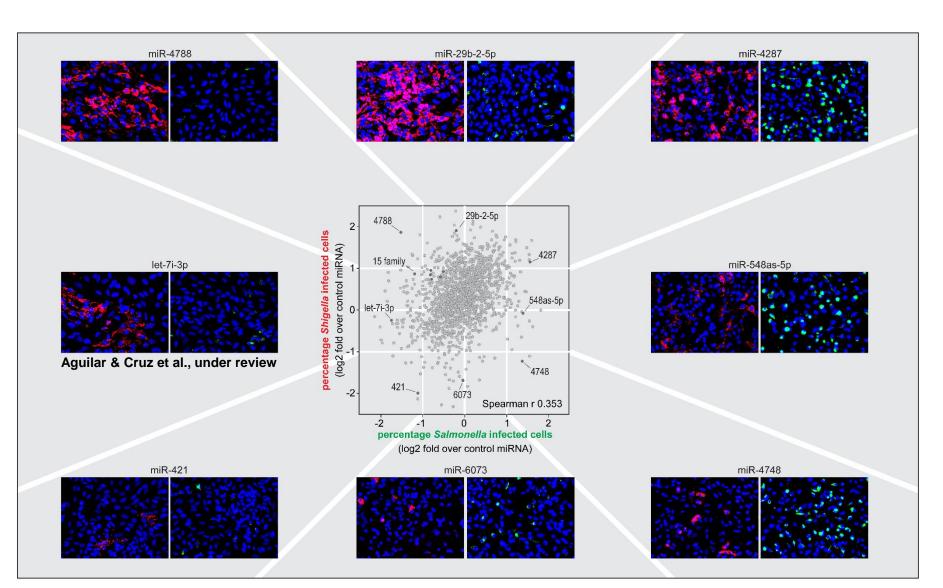
Cruz, Aguilar et al., under review

miRNAs interfere with different stages of *Salmonella* and *Shigella* infection



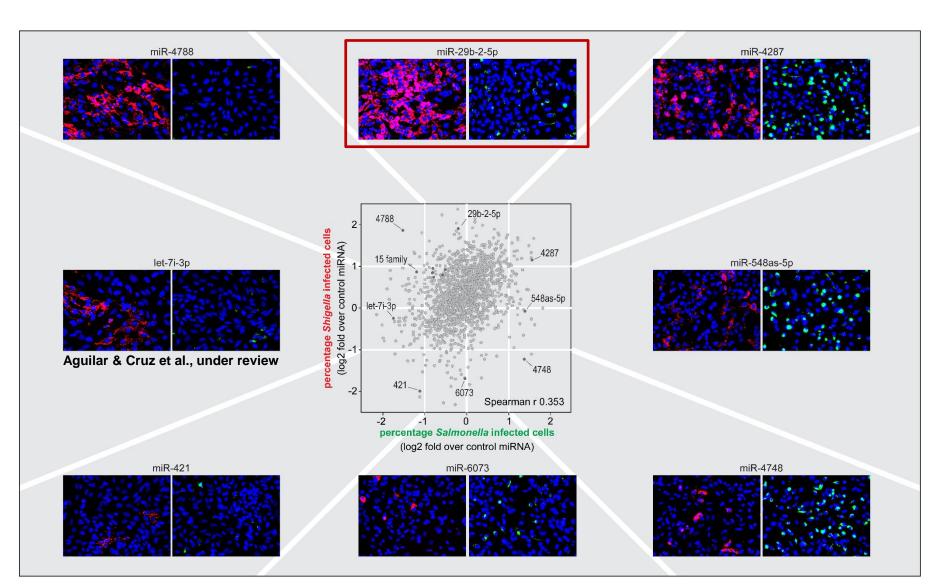
Cruz, Aguilar et al., under review

Salmonella and Shigella infection are regulated by a different set of miRNAs

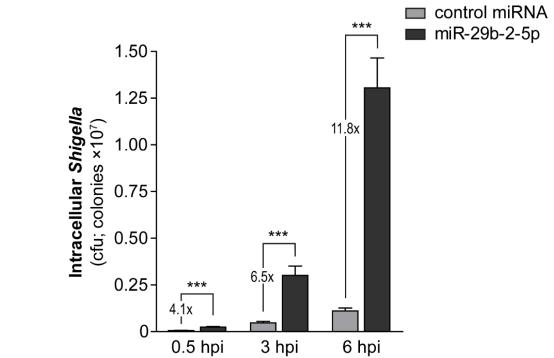


Cruz, Aguilar et al., under review

Salmonella and Shigella infection are regulated by a different set of miRNAs



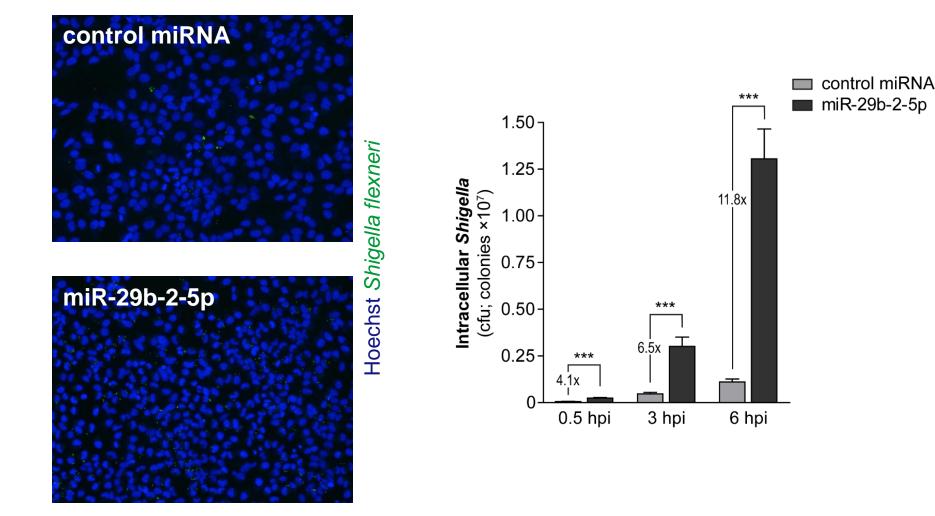
miR-29b-2-5p increases both early and late steps of *Shigella* infection



Hoechst Shigella flexneri

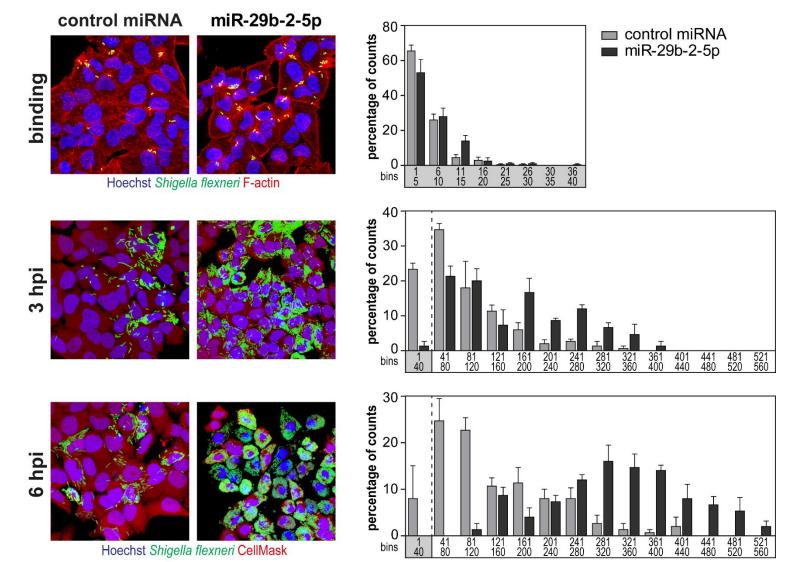
6 h time course

miR-29b-2-5p increases both early and late steps of *Shigella* infection



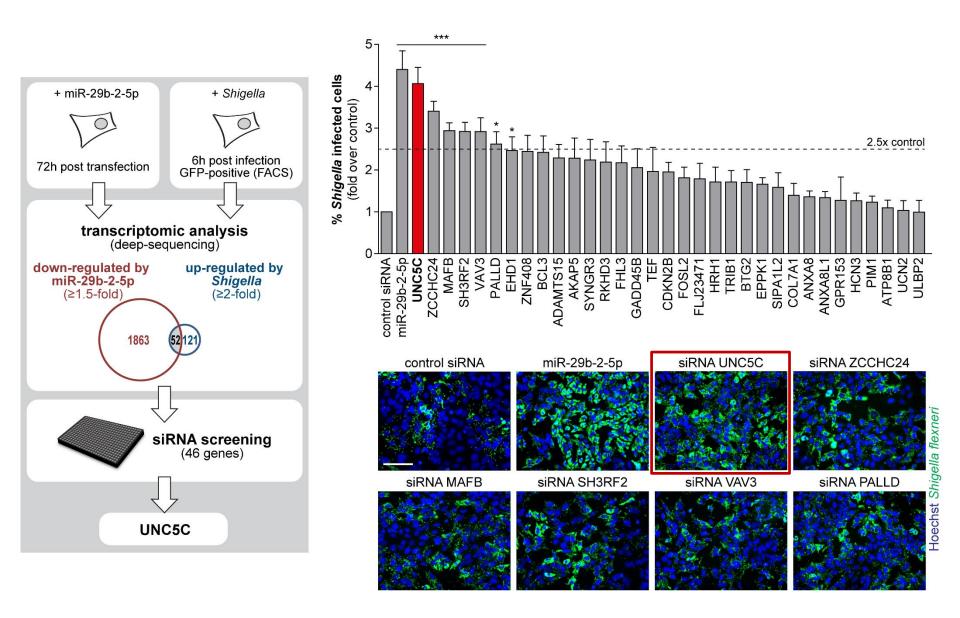
6 h time course

miR-29b-2-5p increases both Shigella binding to host cells and intracellular replication

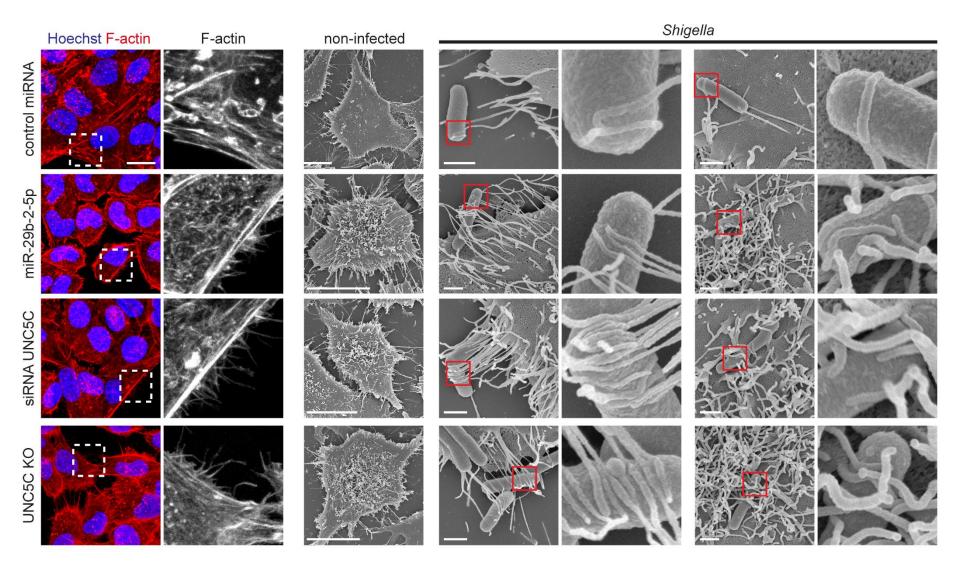


Shigella number/cell (Shigella + cells)

Identification of miR-29b-2-5p targets relevant for *Shigella* infection



miR-29b-2-5p increases filopodia formation and Shigella capture by targeting UNC5C



Sunkavalli, Aguilar et al., PLOS Pathogens (2017) 3(4):e1006327

(slides removed, video cut)

Shedding light on microRNA function via microscopy-based screening

Ines Rodrigues Lopes^{a,b}, Ricardo Jorge Silva^b, Ines Caramelo^{a,b}, Ana Eulalio^{a,*}, Miguel Mano^{b,*}

^a RNA & Infection Group, Center for Neuroscience and Cell Biology (CNC), University of Coimbra, Coimbra, Portugal ^b Functional Genomics and RNA-based Therapeutics Group, Center for Neuroscience and Cell Biology (CNC), University of Coimbra, Coimbra, Portugal

Special Issue on "MicroRNA Biogenesis, Identification, Function and Decay" Methods (2019), 152:55-64

Table 1

Microscopy-based phenotypic screenings using arrayed libraries of synthetic miRNA precursors, mimics and inhibitors.

Phenotype	Library	Species	Cell line	Readout	Reference
Infection					
Salmonella infection	875 human miRNA mimics (Dharmacon)	Human	HeLa	Hoechst, Salmonella-GFP	Maudet et al. (2014) [17]
Shigella infection	2042 human miRNA mimics (Dharmacon)	Human	HeLa	Hoechst, CellMask, Shigella	Sunkavalli et al. (2017) [18]
Flavivirus replication (DENV, WNV, JEV)	1200 human miRNA mimics (Dharmacon)	Human	HeLa	DAPI, Flavivirus envelope	Smith et al. (2017) [19]
Cancer					
Cell proliferation	328 human miRNA precursors (Ambion)	Human	Human primary mammary epithelial cells (HEMC)	DAPI, BrdU	Borgdorff et al. (2010) [20]
Cell migration	904 human miRNA mimics (GenePharma)	Human	HeLa, HepG2, U2OS	area occupied by cells	Zhang et al. (2011) [21]
Cell proliferation and apoptosis	319 human precursor miRNAs (Ambion)	Human	HCT116, LS174T TR4, DLD1 TR7, HT29, Caco2, SW480	Ki-67, cPARP, DAPI	Christensen et al. (2014) [22]
Cell migration	646 human miRNA mimics (GenePharma)	Human	SK-Hep-1	wound healing	Zha et al. (2014) [23]
Apoptosis	895 human miRNA mimics (GenePharma)	Human	HeLa	Hoechst, Annexin V, FITC	Ma et al. (2017) [24]
Fibroblast pro-tumorigenic activity	988 human miRNA mimics (Dharmacon)	Human	Immortalized cancer associated fibroblasts	Hoechst, A549-GFP cells	Andriani et al. (2018) [25]
Cardiovascular					
Cell proliferation	875 human miRNA mimics (Dharmacon)	Rat	Primary neonatal rat cardiomyocytes	Hoechst, α -actinin, Ki67, EdU	Eulalio et al. (2012) [26]
Hypertrophy	230 human miRNA precursors (Ambion)	Rat	Primary neonatal rat cardiomyocytes	DAPI, α-actinin, P4HB	Jentzsch et al. (2012) [27]
SERCA2a expression	875 human miRNA precursors (Ambion)	Human	HEK293	EGFP-SERCA2a-3'UTR	Wahlquist et al. (2014) [28]
Cell proliferation	875 human miRNA mimics (Applied Biosystems)	Human	hiPSC derived cardiomyocytes	DAPI, α -actinin, EdU	Diez-Cuñado et al. (2018) [29]
Cell reprogramming					
iPSC colony formation	379 human miRNA precursors (Ambion)	Mouse	MEF	Oct4-GFP	Pfaff et al. (2011) [30]
Germ layer formation	875 human miRNA precursors (Ambion)	Mouse	mESC	Myh6-EGFP	Colas et al. (2012) [31]
Dedifferentiation of fibroblasts to iPSCs	570 mouse miRNA mimics (Dharmacon)	Mouse	MEF	Oct4-GFP	Judson et al. (2013) [32]
Muscle cell differentiation	870 human miRNA inhibitors (Exiqon)	Human	LHCN	Hoechst, MHC	Polesskaya et al. (2013) [33]
Reprogramming of epithelial cells to organoids	328 human miRNA precursors (Ambion)	Human	Human primary mammary epithelial cells (HEMC)	CD44, CD24, DAPI	Delom et al. (2018) [34]
Others					
Lipid droplet formation	327 human miRNA precursors (Ambion)	Human	Huh7	nuclei, lipid droplets	Whittaker et al. (2010) [35]
Improved protein expression	875 human miRNA mimics (Qiagen)	Human	T-REx-293	Hoechst, NTSR1-GFP	Xiao et al. (2015) [36]
Low-density lipoprotein receptor pathway activity	1719 miRNA mimics (mirVana)	Human	Huh7	Hoechst, DiI-LDL	Goedeke et al. (2015) [37]

Table 3High-content microscopy screening platforms.

Manufacturer/Model	Camera Technology, Bit-depth, pixel number, FOV@ $10 \times^*$	Confocal	Illumination	Autofocus	Transmitted light	Special hardware features				
<i>Acquifer</i> Imaging Machine	https://www.acquifer.de/screening/ sCMOS 16-bit, 4.2 MP 1.8 mm ²	по	LED	laser + image	yes	static sample holder, moving optics block				
GE Healthcare	https://www.gelifesciences.com/en/au/s	shop/cell-imaging-and-ana	lysis/high-conter	nt-analysis-system	s/instruments					
IN Cell Analyzer 2200	sCMOS 16-bit, 5.5 MP, 1.3 mm ²	no	LED	laser + image	yes	optional O_2 control				
IN Cell Analyzer 2500 HS	sCMOS 16-bit, 4.2 MP, 1.3 mm ²	no	LED	laser + image	yes	optional O ₂ control				
IN Cell Analyzer 6000	sCMOS 16-bit, 5.5 MP, 1.3 mm ²	IRIS line scanning	laser	laser + image	yes	optional O ₂ control				
IN Cell Analyzer 6500 HS	sCMOS 16-bit, 4.2 MP, 1.3 mm ²	IRIS/EDGE line scanning	laser	laser + image	yes	optional O ₂ control				
Molecular Devices https://www.moleculardevices.com/products/cellular-imaging-systems#High-Content-Imaging										
ImageXpress Micro 4	sCMOS 16-bit, 4.6 MP, 2.0 mm ²	optional	LED	laser + image	optional	optional on-board liquid handling				
ImageXpress Micro Confocal	sCMOS 16-bit, 4.2 MP, 2.0 mm ²	AgileOptix spinning disk	LED	laser + image	optional	optional on-board liquid handling				
Perkin Elmer	Perkin Elmer http://www.perkinelmer.com/category/high-content-screening-instruments-microscopes									
Operetta CLS	sCMOS 16-bit, 4.6 MP, 1.7 mm ²	optional	LED	laser	yes	automated water immersion objectives				
Opera Phenix	sCMOS 16-bit, 4.6 MP, 1.7 mm ²	Nipkow spinning disk	laser	laser	yes	automated water immersion objectives; up to 4 cameras				
ThermoFisher Scientific	https://www.thermofisher.com/pt/en/h	ome/life-science/cell-analy	/sis/cellular-imag	ging/high-content-	screening/high-conten	at-screening-instruments.html				
CellInsight CX5	CCD 14-bit, 4.9 MP, 1.0 mm ²	no	LED	Image	yes	, i i i i i i i i i i i i i i i i i i i				
CellInsight CX7	CCD 14-bit, 4.9 MP, 1.0 mm ²	CrEST spinning disk	LED	laser + image	yes					
CellInsight CX7 LZR	CCD 14-bit, 4.9 MP, 1.0 mm ²	CrEST spinning disk	laser	laser + image	yes					
Yokogawa	https://www.yokogawa.com/solutions/p	products-platforms/life-scie	ence/high-conten	t-analysis/						
CQ1	sCMOS 16-bit, 5.5 MP, 2.4 mm ²	Nipkow spinning disk	laser	laser + image	optional					
CellVoyager CV8000	sCMOS 16-bit, 4.0 MP, 1.7 mm ²	Yokogawa spinning disk CSU	laser	laser + image	yes	automated water immersion objectives; up to 4 cameras; built-in stage incubator for live cell imaging				
Zeiss	https://www.zeiss.com/microscopy/int/	products/imaging-systems.	html							
CellDiscoverer 7	CCD 14-bit, 6 MP, 0.6 mm ²	no	LED	laser + image	yes	automated water immersion objectives; up to 2 cameras; optional liquid handling; 12 MP external camera				

* FOV@10 \times – Field of view with a 10 \times objective.

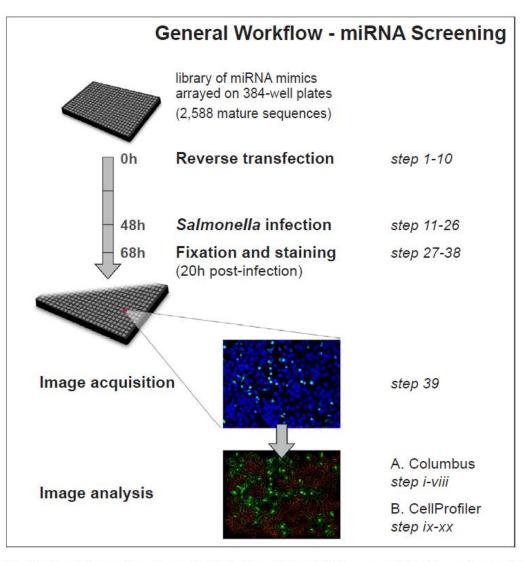


Fig. 1. Overview of a general workflow for miRNA screening. Procedures for reverse transfection, *Salmonella* infection and cell fixation and staining, as well as image acquisition and analysis using a commercial and an open-source image analysis software platform are detailed in Section 3.

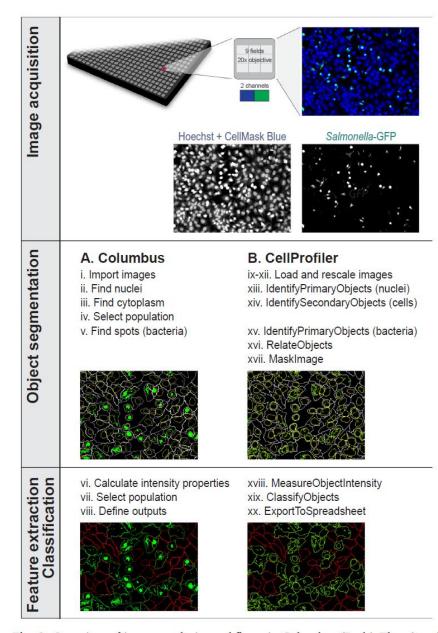


Fig. 3. Overview of image analysis workflows in Columbus (PerkinElmer) and CellProfiler (open-source) image analysis software platforms. Details of each step related to object segmentation, feature extraction and classification are provided in Section 3.5.

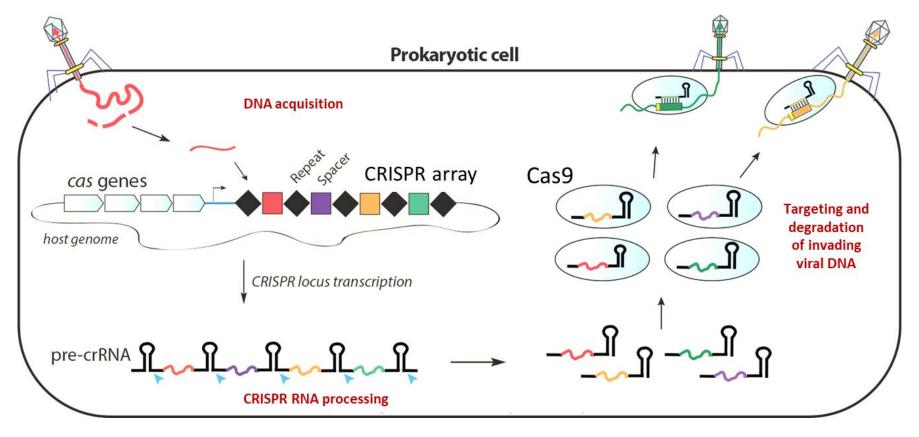
CRISPR/Cas9 technology

Precise genome editing and beyond

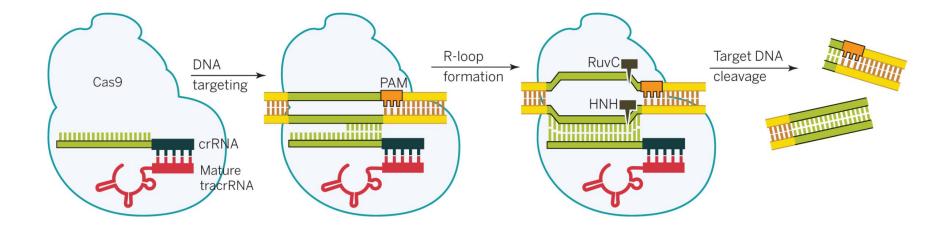
The CRISPR/Cas System

Bacterial and archea adaptive

immunity <u>Clustered Regularly Interspaced Short Palindromic Repeats / CRISPR-</u> associated proteins



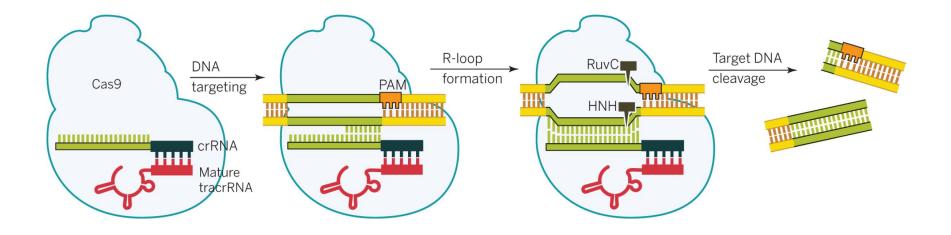
The CRISPR/Cas System

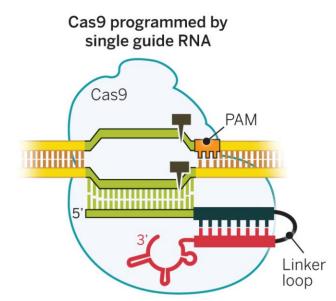


CRISPR (clustered regularly interspaced palindromic repeats)

Cas (CRISPR-associated proteins).

The CRISPR/Cas System

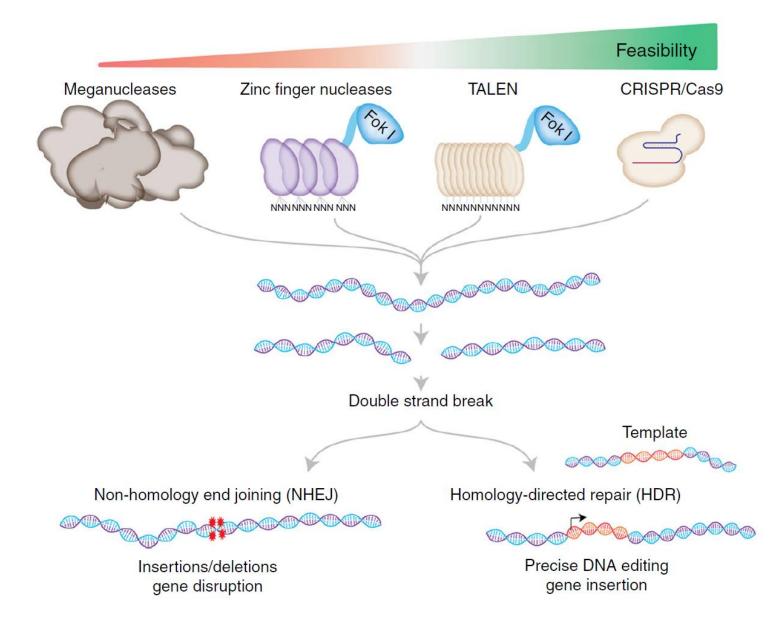




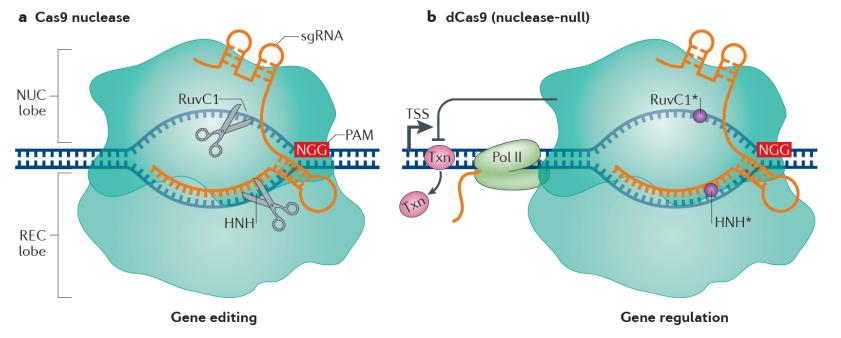
CRISPR (clustered regularly interspaced palindromic repeats)

Cas (CRISPR-associated proteins).

Nuclease-induced genome editing



CRISPR/Cas system: beyond cutting

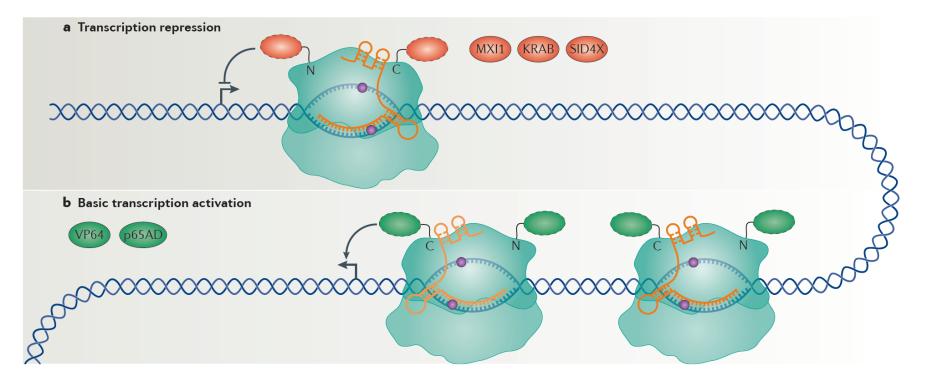


dCas9 maintains DNA targeting specificity, without endonuclease activity

(Cas9 with nickase activity induces single-strand DNA cuts)

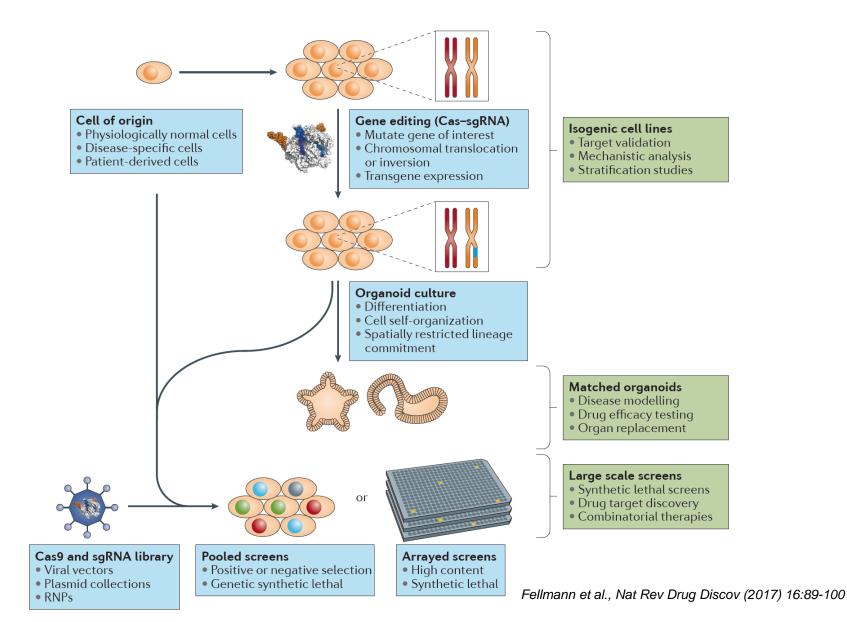
CRISPR/Cas system for gene regulation

CRISPRi

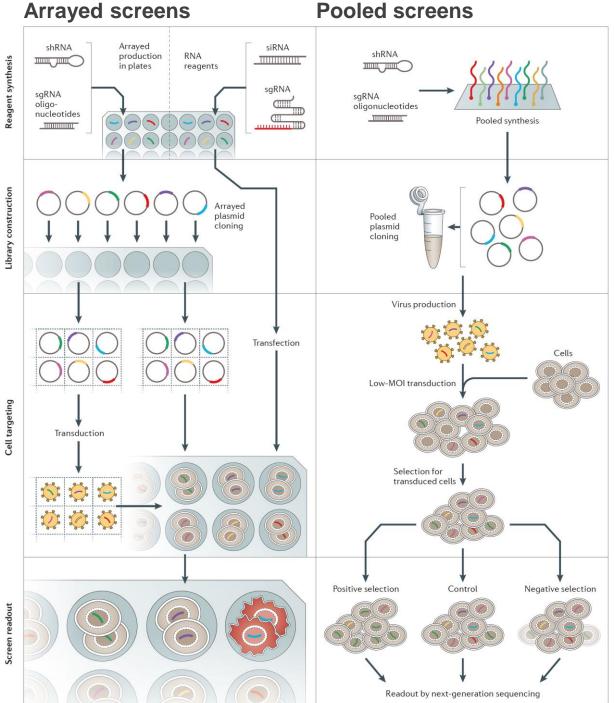




CRISPR/Cas for disease modelling and large-scale screening



Screening approaches



Shalem et al., Nat Rev Genet (2015) 16:299-311



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