

# Three-dimensional cellular spheroides in oncology research- a bridge between *in vitro* and *in vivo* studies

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# Spheroids

Discovery

Validation, testing

**IN VITRO**  
Primary screening  
(Cell based assays)

**IN VIVO**  
Secondary screening  
(Animal models)

**CLINICAL STUDIES**  
Secondary screening  
(Patient)

## SPHEROID

- 3D spheroid cultures better represent *in vivo* tissues
- more realistic cell-cell and cell-matrix interactions
- nutrient, oxygen and waste gradients present
- increases confidence in *in vitro* testing results

## Targeted research areas:

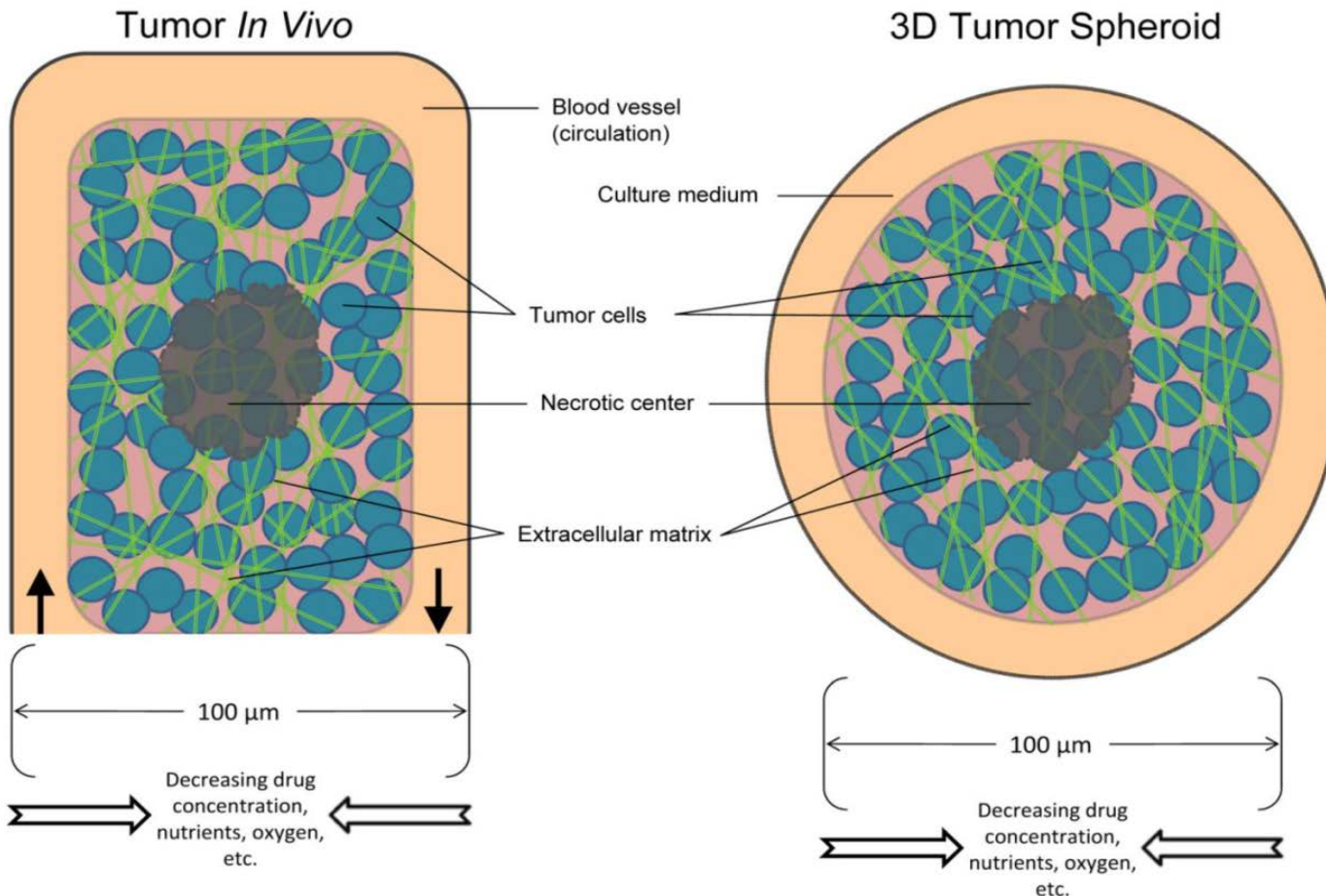
- cancer research (model for the microareas in the tumor, growth, metastasizing, invasion, angiogenesis)
- screening for new drugs (high throughput screening)
- toxicology
- stem cell research
- tissue engineering

## Characteristics of 2D and 3D cell cultures-spheroids

Characteristic	2D	3D
Secretion of extracellular matrix (ECM)	no	collagens, laminin, fibronectin, glycosaminoglycans; similarly organized as in the tissues
Cell-cell interactions	no	yes (homo and hetero interactions)
Cell- ECM interactions	no	yes (cell-matrix adhesions)
Proliferation and growth rate	activated, fast	regulated; slower
Gene expression and protein secretion profile	qualitatively and quantitatively different	upregulation of the expression of genes: involved in progression and metastatic processes (IL-8, GRO $\alpha$ ali MIP-3 $\alpha$ ), regulation of the ECM components, intercellular junctions secretion of: growth factors, proangiogenic factor VEGF, TNF $\alpha$
Drug sensitivity	more	less

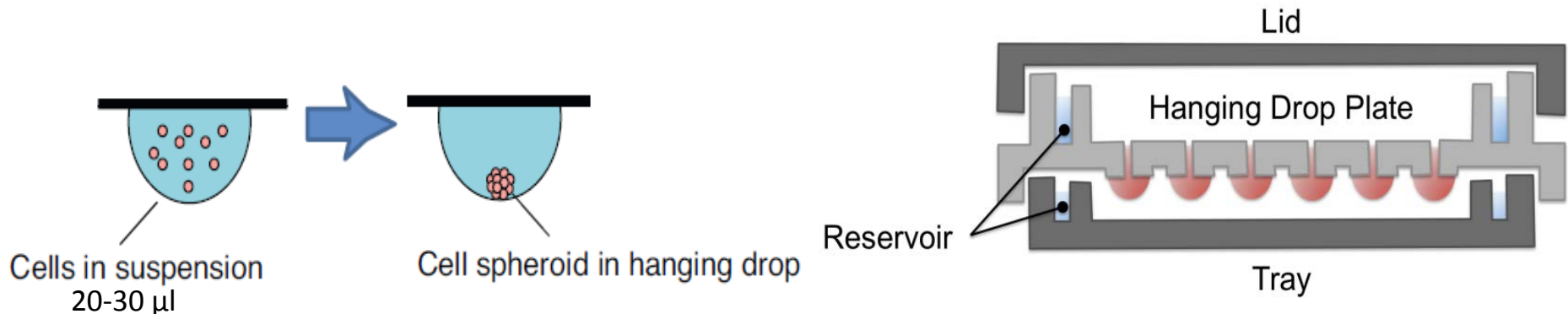
# Characteristics of spheroides

- nutrient, oxygen and waste gradients
- necrosis area  $2r > 100 \mu\text{m}$ ;  $2r > 500 \mu\text{m}$  hipoxia and necrosis in the center
- transplation *in vivo*



## 3D cell culture-spheroid methods

Method	Advantages	Disadvantages
Hanging drop	<ul style="list-style-type: none"> <li>-inexpensive (96 well plate, petry dish coated with agar)</li> <li>-homogenous</li> <li>-suitable for high-throughput testing</li> <li>-easily accessible spheroides</li> </ul>	<ul style="list-style-type: none"> <li>-expensive if using specialised plates</li> <li>-difficult exchange of medium due to small culture volume</li> <li>-labour intensive if preparing the plates</li> <li>-diameter up to 600 <math>\mu\text{m}</math></li> </ul>



# Hanging drop: Human adenocarcinoma colon cancer HT29 spheroides

3 days after seeding cells

300 cells

500 cells

1000 cells

2000 cells

3000 cells



10 days after seeding cells

300 cells

500 cells

1000 cells

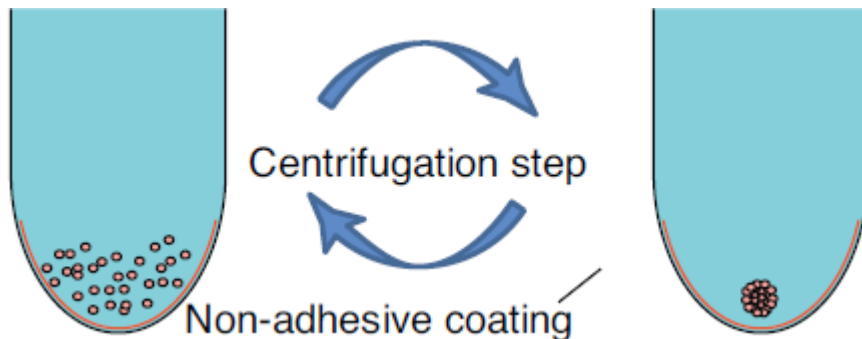
2000 cells

3000 cells

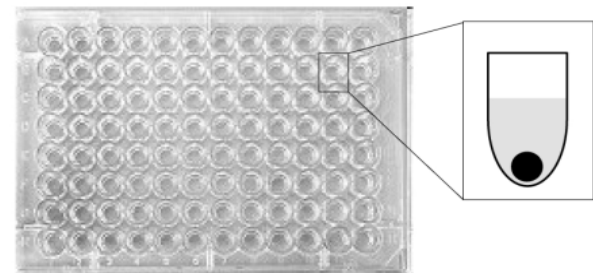


# 3D cell culture-spheroid methods

Method	Advantages	Disadvantages
Forced floating	<ul style="list-style-type: none"> <li>-simple, fast (96 well plate ULA or normal plate coated with agar /different biopolymer; round or conical bottom)</li> <li>-inexpensive</li> <li>-suitable for high-throughput testing</li> <li>-easily accessible spheroides</li> <li>-more culture medium</li> <li>-diameter up to 1500 <math>\mu\text{m}</math></li> </ul>	<ul style="list-style-type: none"> <li>-variability in size and shape (fixed cell no./well!)</li> <li>-plate coating is relatively labour intensive</li> </ul>



## Spheroid generation



ULA 96-well round-bottom plate

# Forced floating: Human squamous cell carcinoma FaDu spheroides

3 days after seeding cells on 96 well plate coated with agar

250 cells

500 cells

1000 cells

2000 cells

3000 cells



2r 167,4µm ± 4,0

249,9µm ± 16,1

279,6µm ± 23,8

369,1µm ± 13,4

416,6µm ± 9,7

13 days after seeding cells on 96 well plate coated with agar

250 cells

500 cells

1000 cells

2000 cells

3000 cells



2r 667,6µm ± 10,3

846,8µm ± 17,5

989,9µm ± 32,1

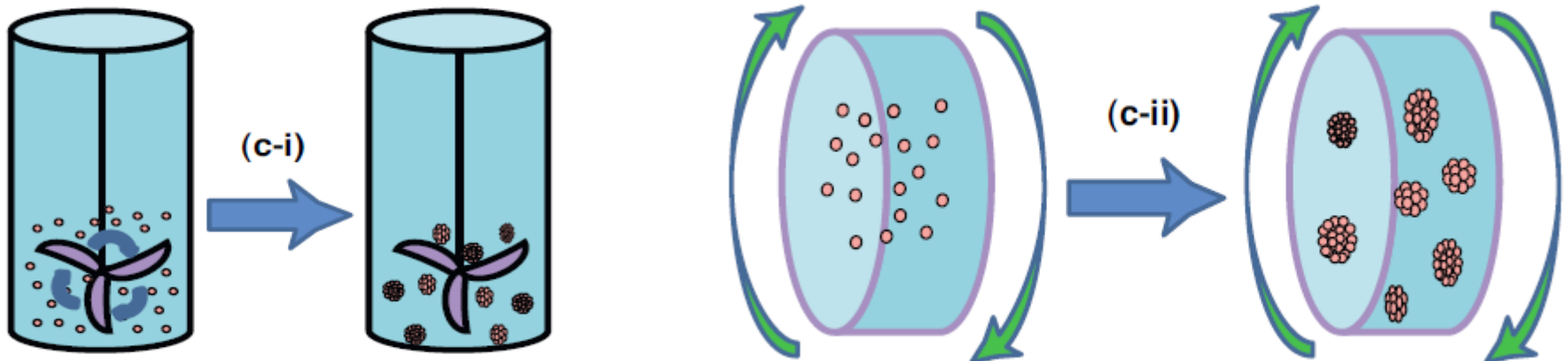
1085,4µm ± 120,5

1073,8µm ± 47,9



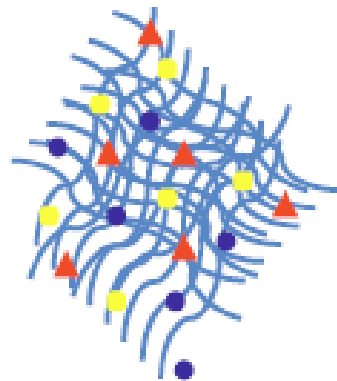
# 3D cell culture-spheroid methods


Method	Advantages	Disadvantages
Agitation-based approaches: -spinner flask bioreactors -rotational culture system	-simple to culture cells -large scale production relatively easily achievable -motion of culture assists nutrient transport -easily accesible spheroides -diameter up to 1 cm	-specialised equipment -no control over size; additional culture step needed to uniform the size -time consuming due to extra step required for homogenous spheroid -cells exposed possibly to shear force in spinner flask (? sensitive cells)



# 3D cell culture-spheroid methods

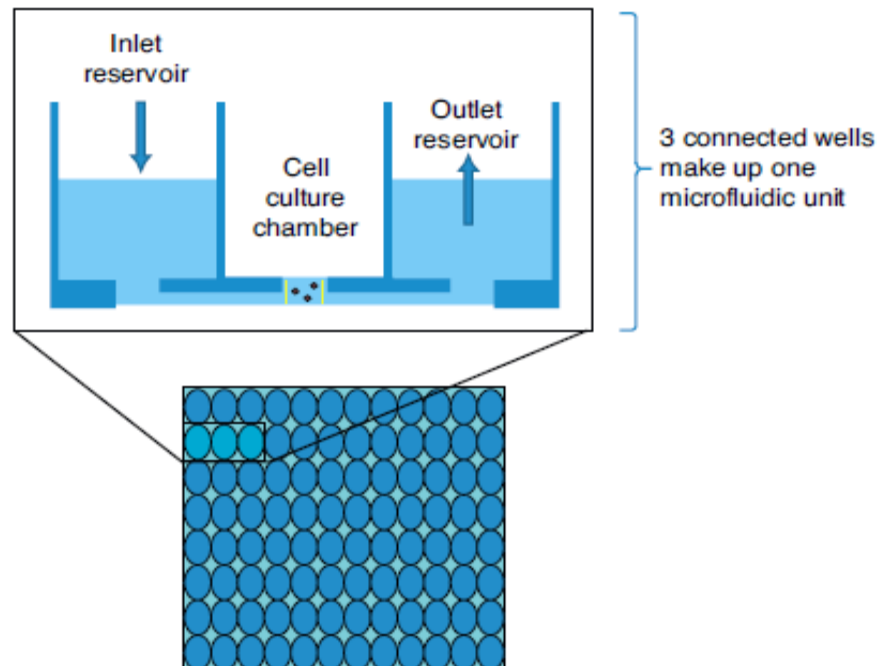
Method	Advantages	Disadvantages
Matrices (matrigel) and scaffolds (collagen, laminin, alginate and other biodegradable materials, which form hydrogels)	<ul style="list-style-type: none"><li>-provide 3D support that mimics in vivo</li><li>-some incorporate growth factors</li></ul>	<ul style="list-style-type: none"><li>-expensive for large scale production</li><li>-difficulty in retrieving cells following 3D culture formation</li><li>-nonhomogenous size</li></ul>



 - Growth factors

# 3D cell culture-spheroid methods

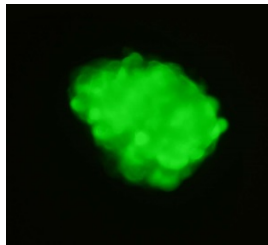
Method	Advantages	Disadvantages
Microfluidic cell culture platforms	-suitable for high throughput testing	-require specialised equipment -further analysis of spheroides are difficult



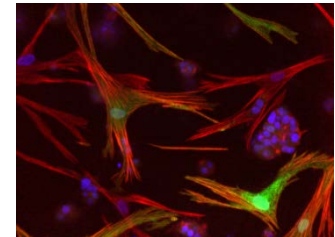
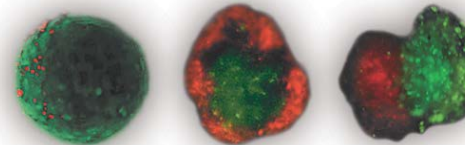
# Spheroides-cocultures

- stable transfected cells with reporter gene (GFP, Luc, dsRed,  $\beta$ -gal) or labeled with fluorescent dye enable easy monitoring and analysing

HepG2-GFP,  
10 days old

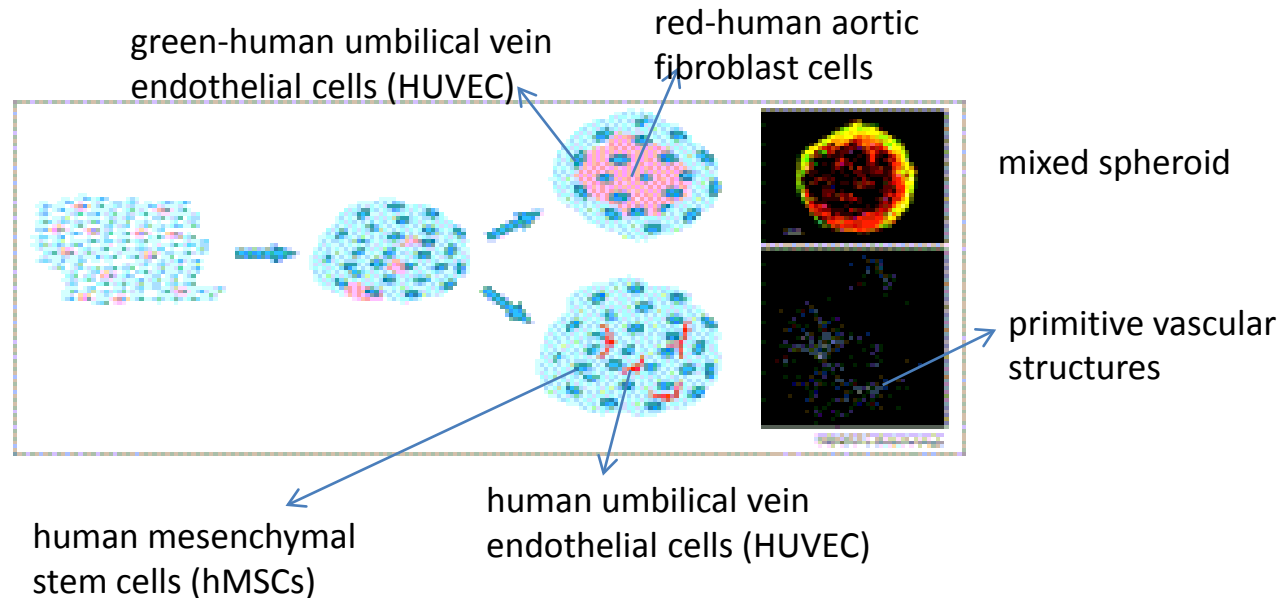


- fibroblasts with tumor cells (epithelial mesenchimal transition-EMT)



## Model system for vascularization

- normal endothelial cells with fibroblasts or stem cells (vascularization)

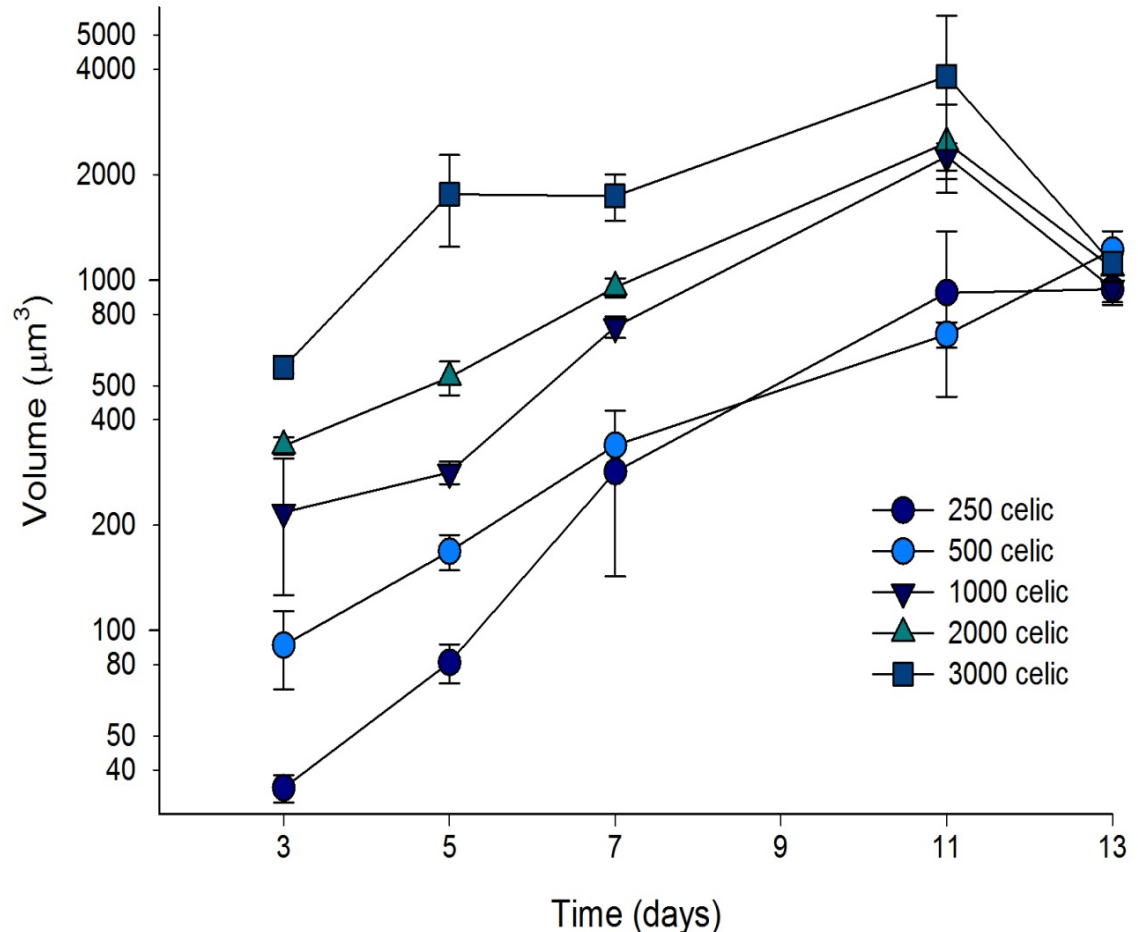


# Analysing and evaluating of spheroides

## Growth analysis:

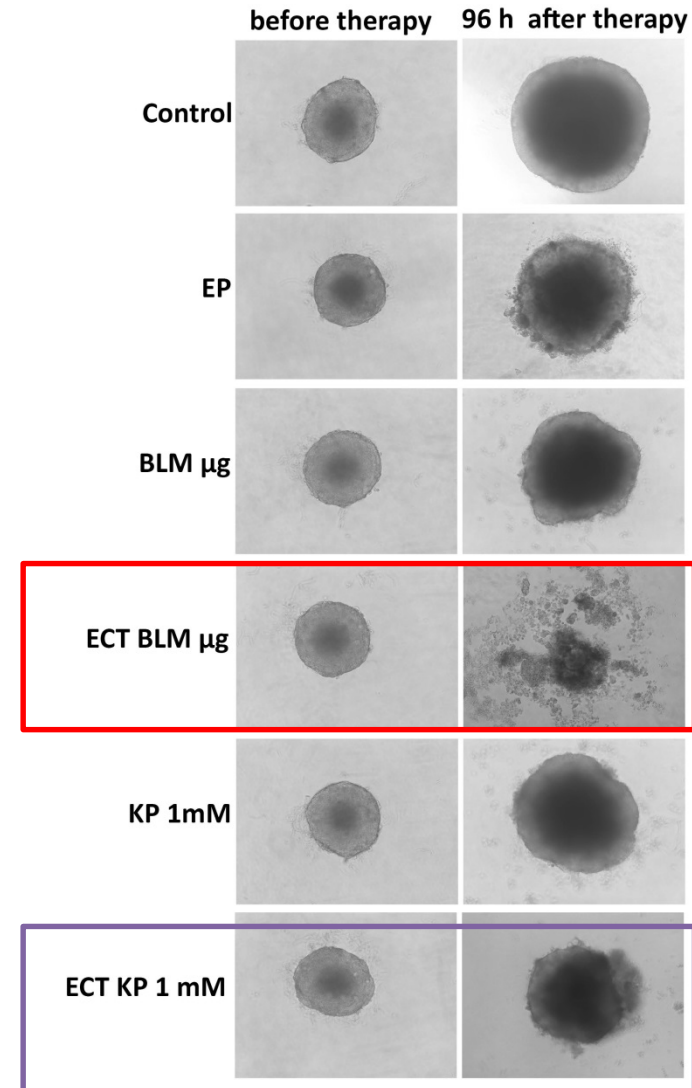
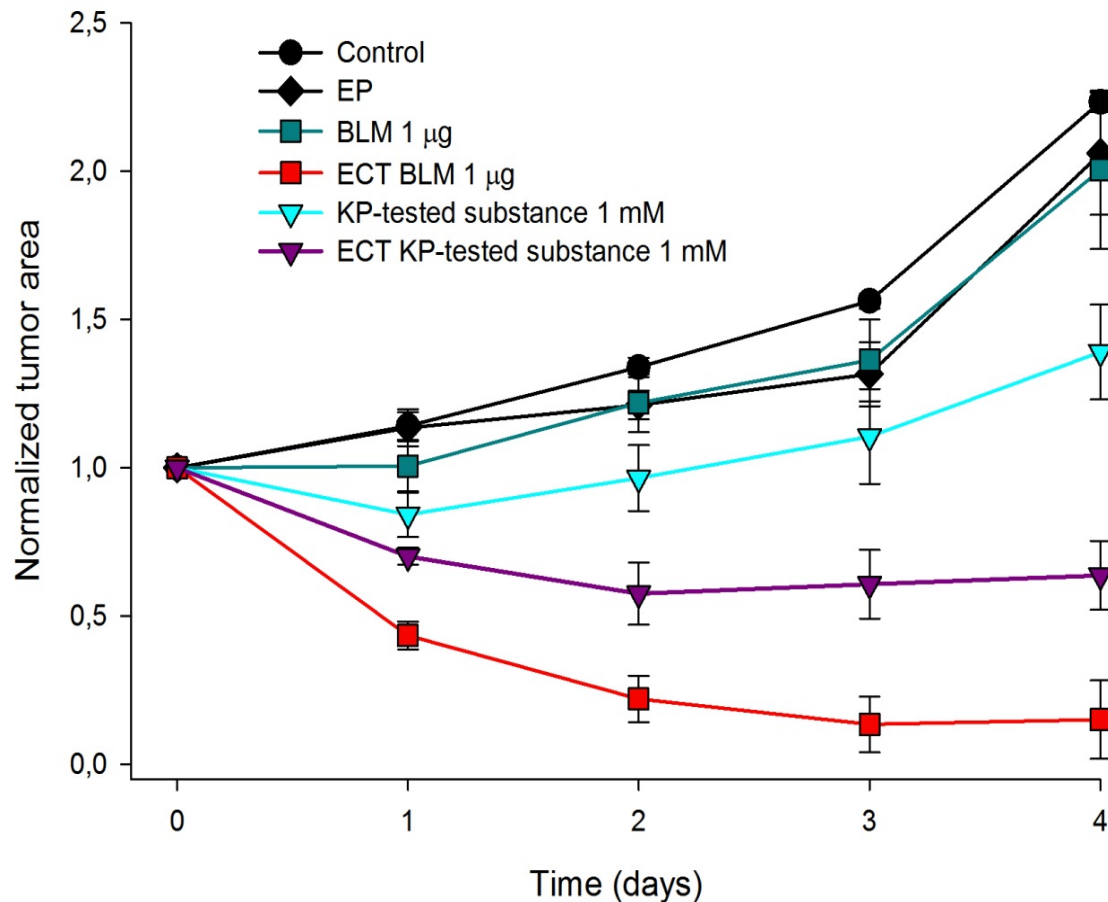
- inverted light or fluorescent microscopy (software for measurement)
- microplate reader (Tecan; Presto Blue, Alamar Blue, MTS) for evaluating viability, proliferation, migration
- cytometry (Celigo cytometer, fast& automated analysing)
- confocal microscopy

Growth of human squamous cell carcinoma FaDu spheroides



# Analysing and evaluating of spheroides after electrochemotherapy

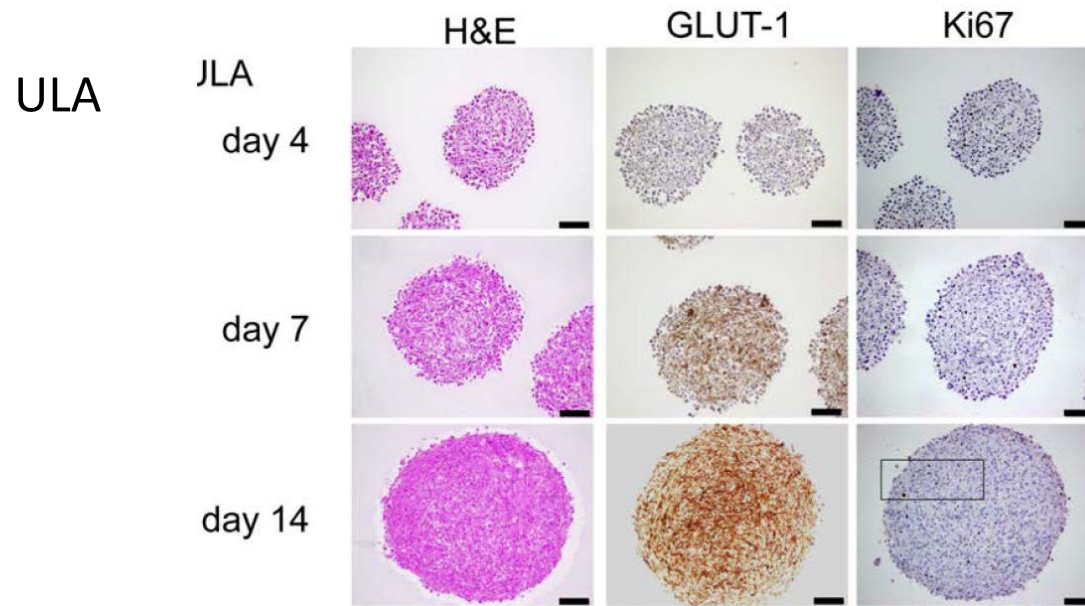
- electrochemotherapy and testing of new drugs (colon cancer carcinoma HT 29spheroides)



# Analysing and evaluating of spheroides

Hystological analysis of spheroides:

- embeeding in parafin, Tissue Tek
- immunohistochemistry analysis: hematoxylin and eosin staining, gradient of proliferation (Ki67), hypoxia (glucose transporter 1; GLUT-1)



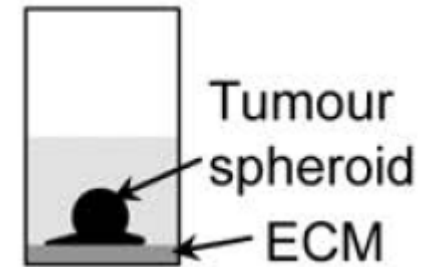
Vinci et al., 2012, BMC Biology, 10:29

Standard analysis on molecular and protein level

# Analysing and evaluating of spheroides

## Migration assay on matrix protein:

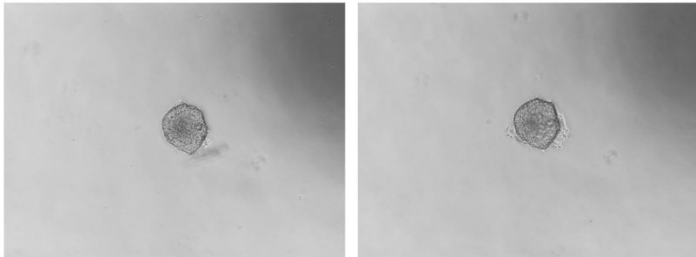
- migration of cells from spheroid on matrix proteins (fibronectin)



### Inverted microscopy

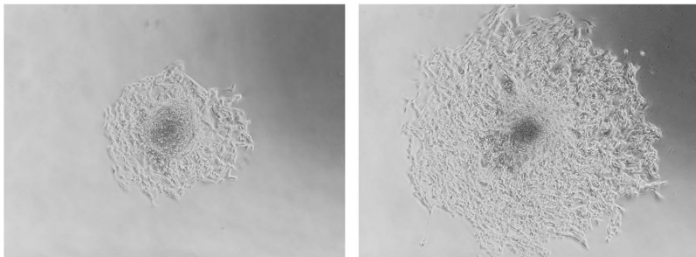
4 h

8 h



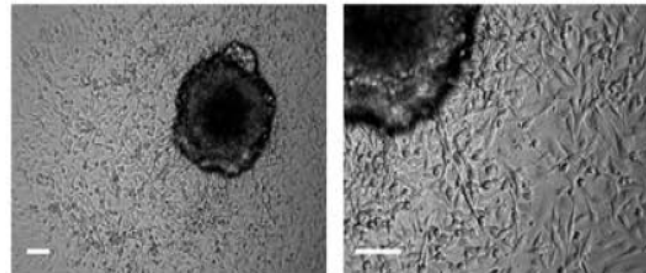
24 h

48 h

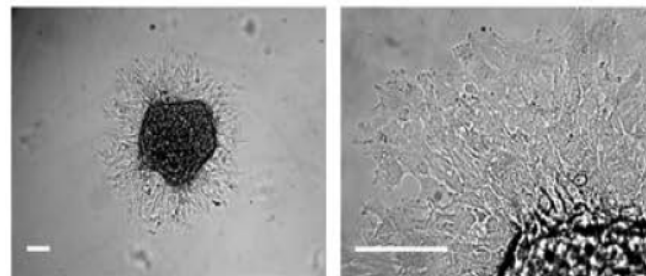


Mouse mamary carcinoma TSA spheroides

### Celigo™ cytometer



U-87 MG  
dispersed migration



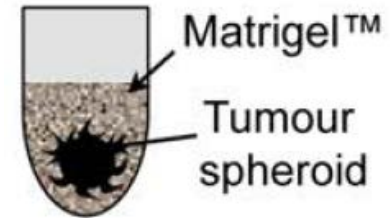
KNS42  
radial migration



# Analysing and evaluating of spheroides

## Matrigel invasion assay :

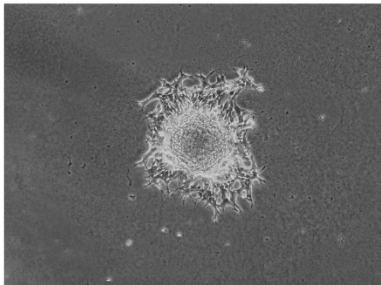
- tumor spheroid embedded in Matrigel



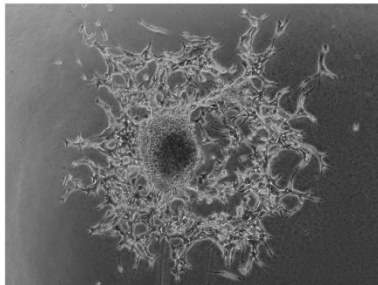
Inverted microscopy

Mouse mamary carcinoma TSA spheroides

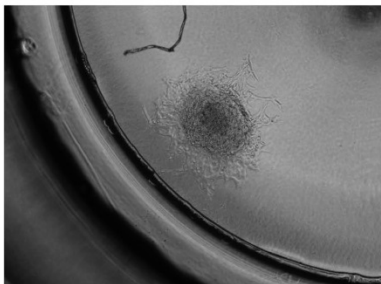
24 h



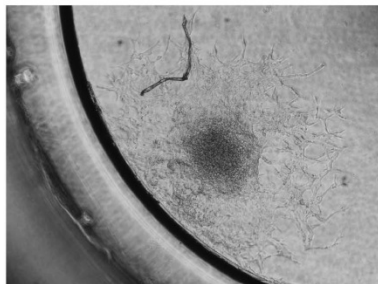
48 h



24 h

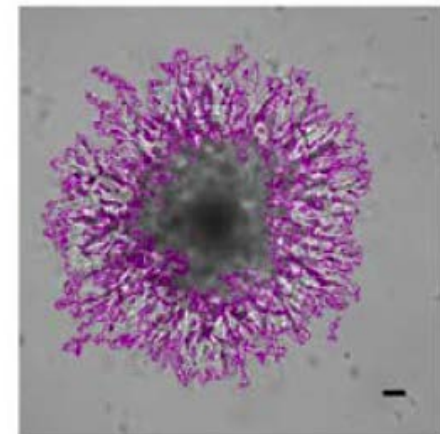
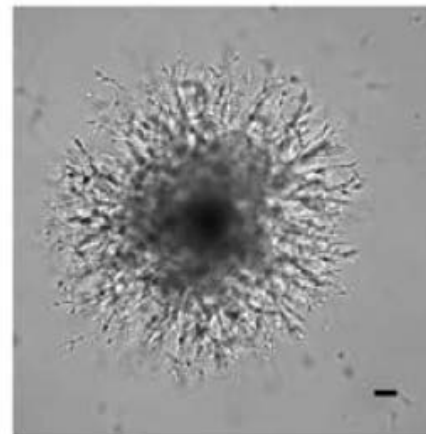


48 h



Celigo™ cytometer

U-87 MG tumour spheroid invasion



# Conclusions

- simple and quite unexpensive to grow the spheroides
- the interactions and communications between the cells in spheroides, secretion of extracelullar matrix - in vivo phenotype of cells is retained
- in spheroides nutrient, oxygen and waste gradients are present
- studies performed on spheroides gives us more **reliable** results *in vitro*
- **importantly complies with the ethical principles of animal research (3 R's: Reduction, Refinement, and Replacement).**

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- LEA EBAM

