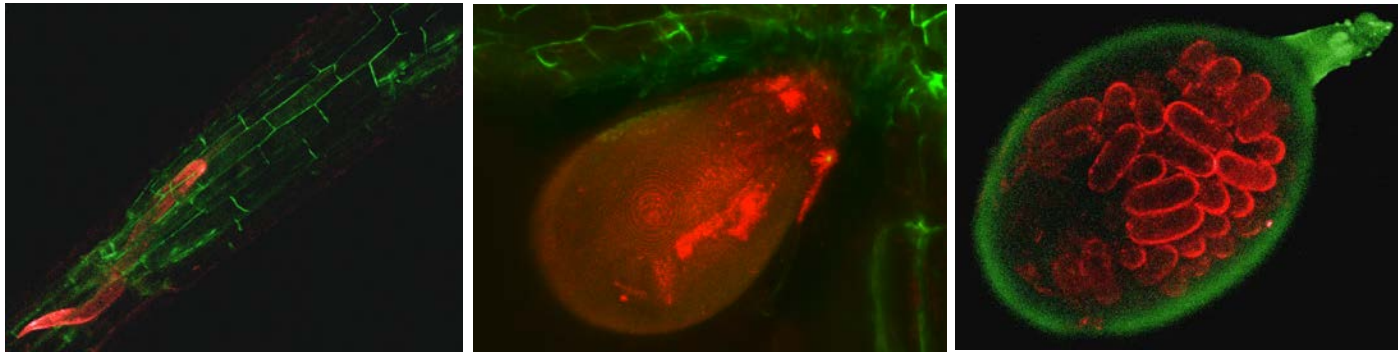


# Nematode effectors – function and resistance

John Jones *et al.*



[john.jones@hutton.ac.uk](mailto:john.jones@hutton.ac.uk)



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**Hutton**  
**Institute**



University of  
**St Andrews**

# Plant parasitic nematodes are damaging pathogens



## Damage valued at \$100 billion pa

- PPNs eat 10% of the world's agricultural output
- 50% of crop losses in developing nations are due to pests and diseases
- Major threat to global food security

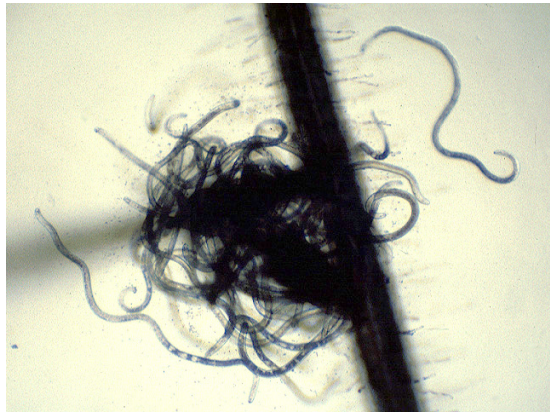


## The most damaging species are root-knot nematodes and cyst nematodes

- Potato cyst nematodes are a major problem in the UK
- Effective nematicides being withdrawn

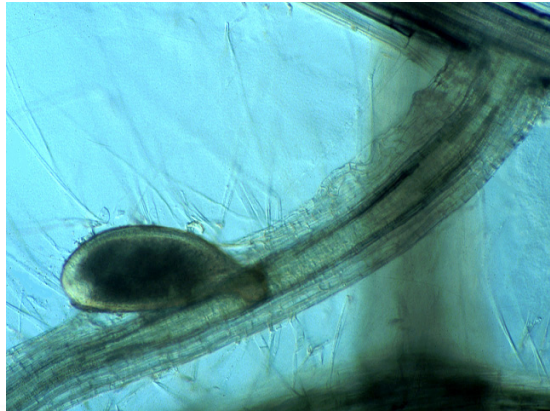
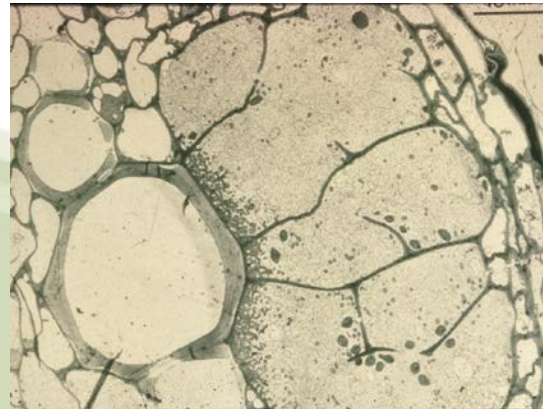


# Plant parasitic nematodes can be biotrophic pathogens

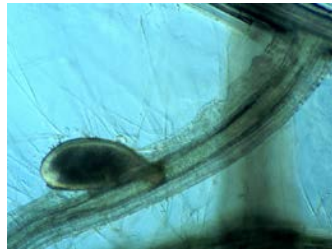
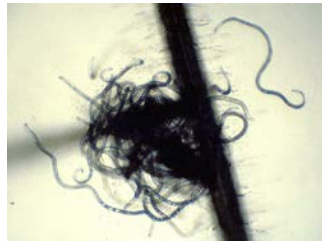


*G. pallida* life cycle

**Syncytium:** kept alive for up to 6 weeks



# *G. pallida* genome and transcriptome resources



Full genome  
sequence

-----  
parasitic

-----  
parasitic



# *G. pallida* genome and transcriptome resources

**Males**



**Eggs**



**J2**

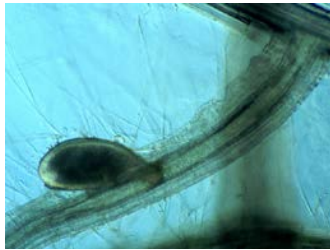


parasitic

**7dpi**



**21dpi**



**28dpi**

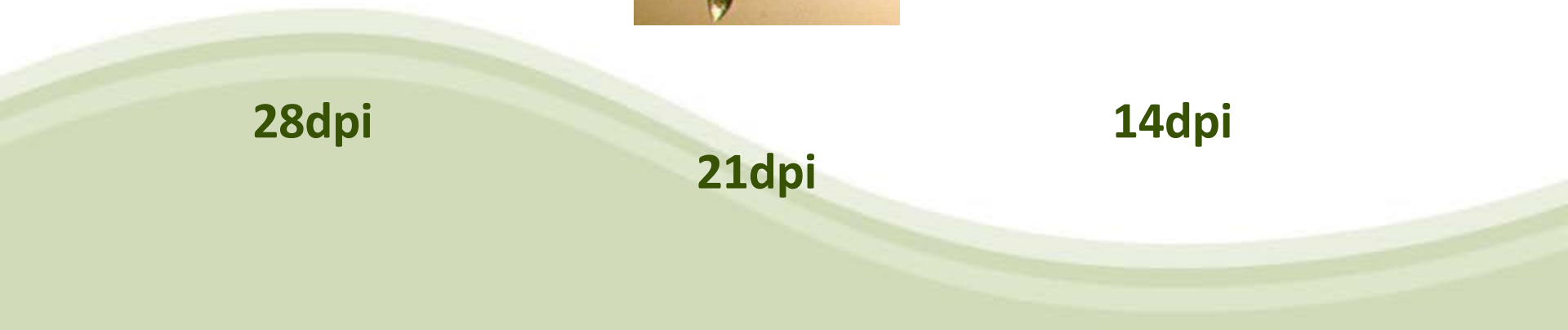


parasitic

**35dpi**



**Full genome  
sequence**



# *G. pallida* genome and transcriptome resources

**Males**



**Eggs**



**J2**

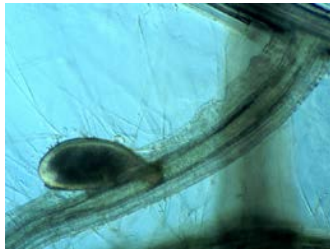


parastic



**7dpi**

**Full genome  
sequence**



**28dpi**

**21dpi**

**14dpi**

parastic

**35dpi**

**Powerful tools for understanding plant nematode biology**

# Effectors of plant parasitic nematodes



## Effectors

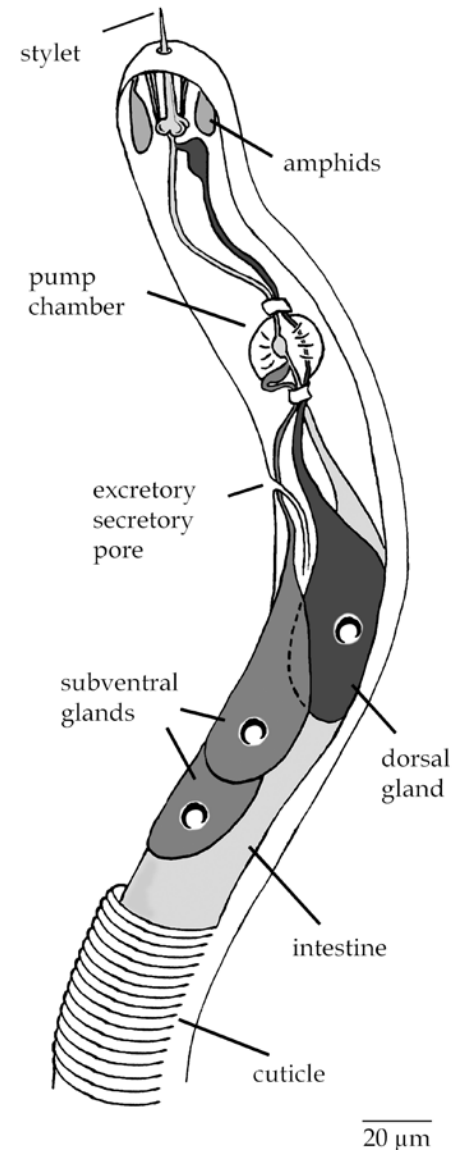
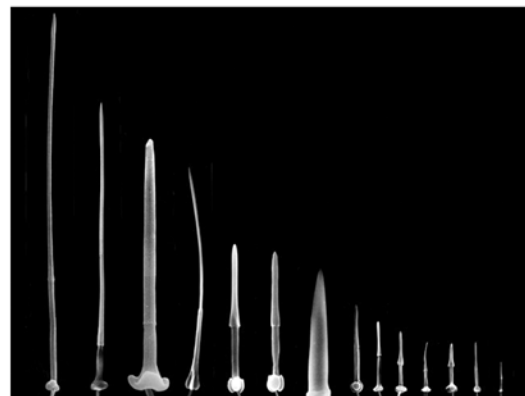
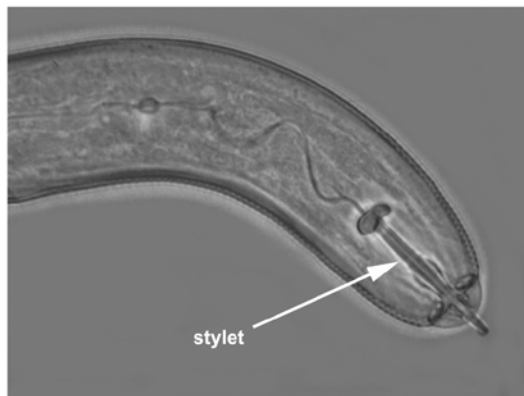
*Proteins secreted by the nematode into the host that manipulate the plant to the benefit of the nematode*

## Effectors play roles in:

- Invasion & migration
- Induction of the feeding site
- Suppression of host defence responses

## Nematode effectors (mainly) from 2 sets of gland cells

- Dorsal gland cell
- Subventral gland cells



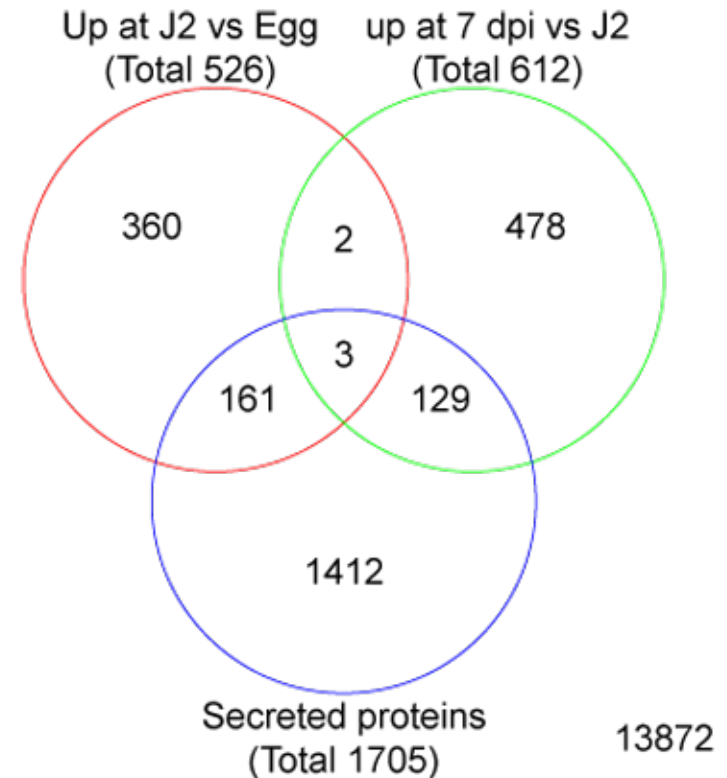
# Identifying effectors from nematodes



**BLAST search *G. pallida* genome with effectors from other nematodes**

**Bioinformatic identification of “novel” effector candidates**

- Signal peptide, no transmembrane domain
- Upregulated in J2 vs egg or parasitic vs J2





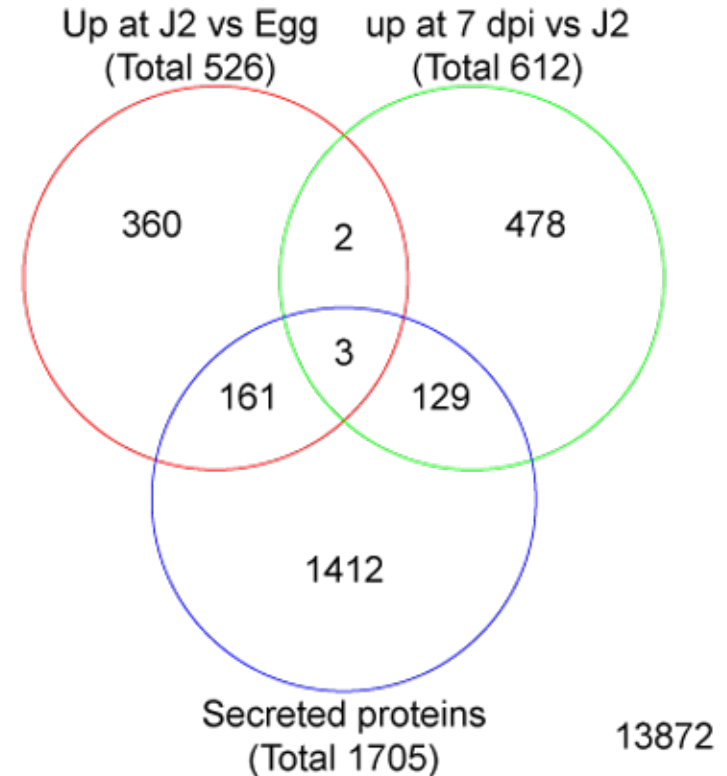
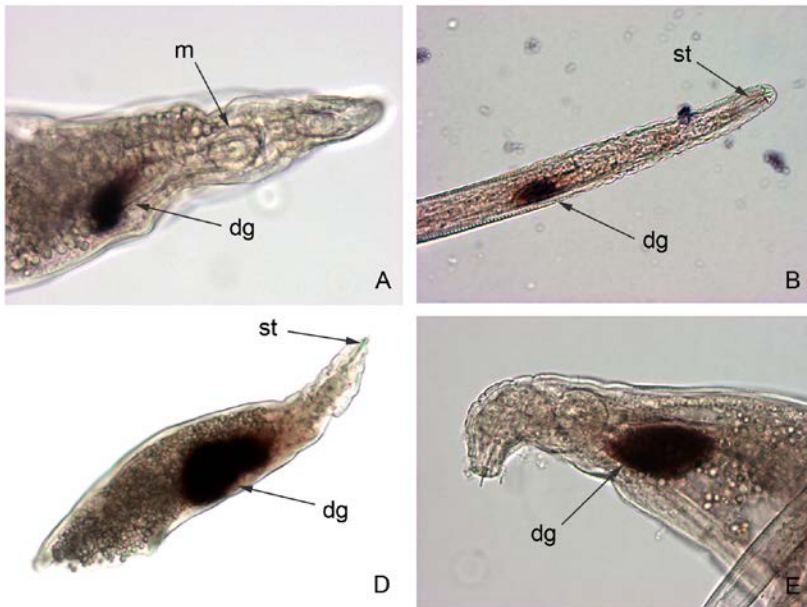
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Bioinformatic identification of “novel” effector candidates

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Confirm by looking at where candidates are expressed in the nematode

# Categorising/prioritising effectors

~ **500 potential effectors identified**

Need to prioritise

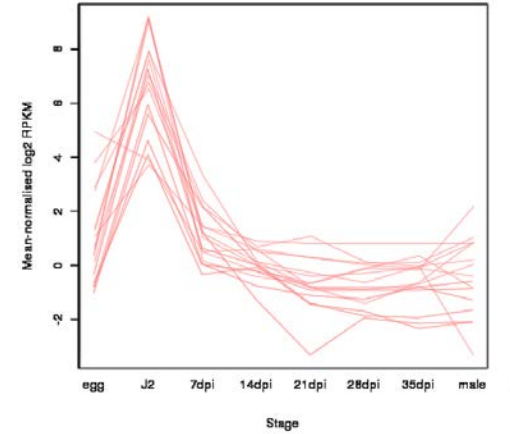
- Presence in a range of populations
- Timing of expression
- RNAi effects
- Subcellular localisation in plants

# Categorising/prioritising effectors

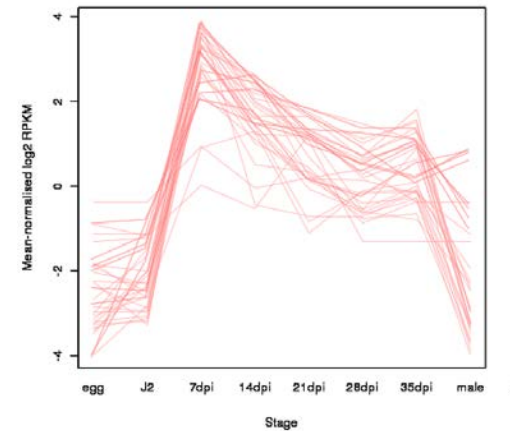
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**J2 specific**



**Parasitic stages**



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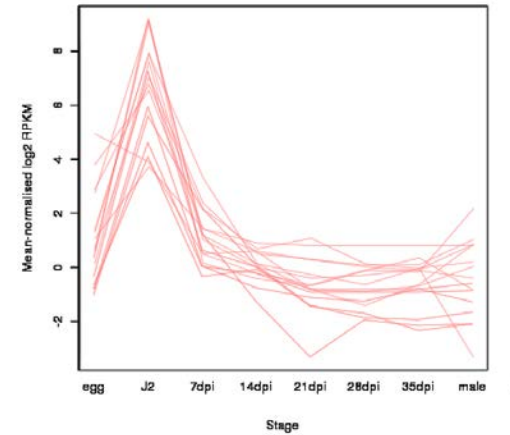
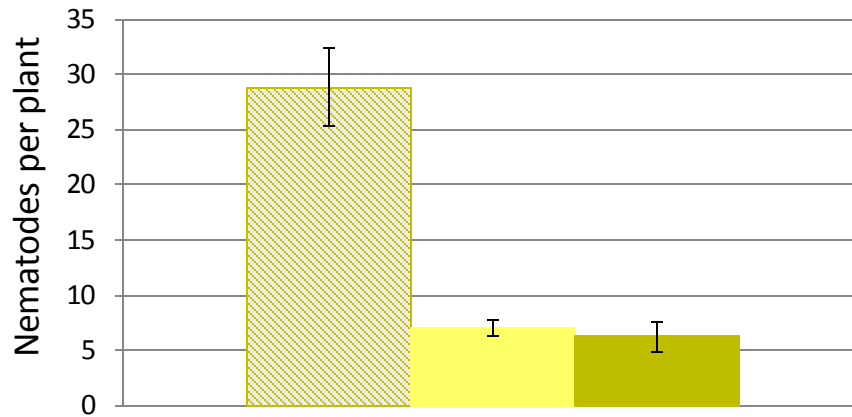
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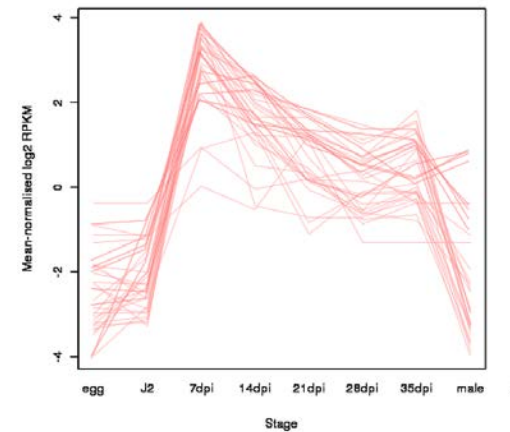
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Control    Knockout line 1    Knockout line 2



J2 specific



Parasitic stages



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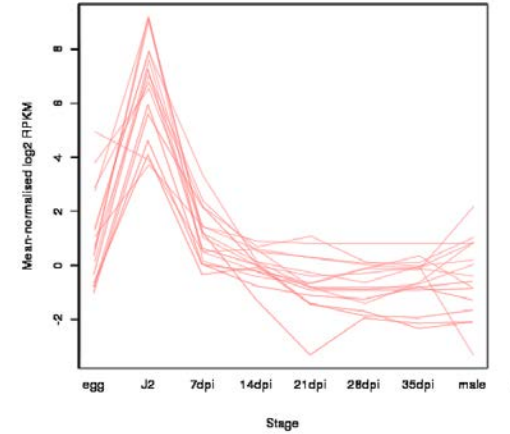
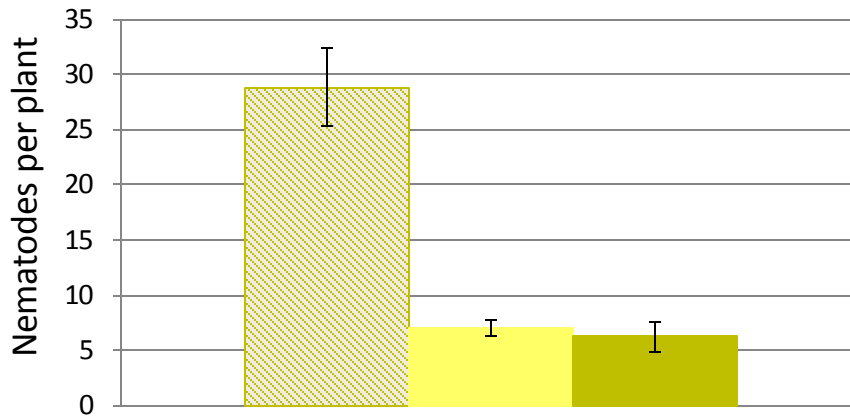
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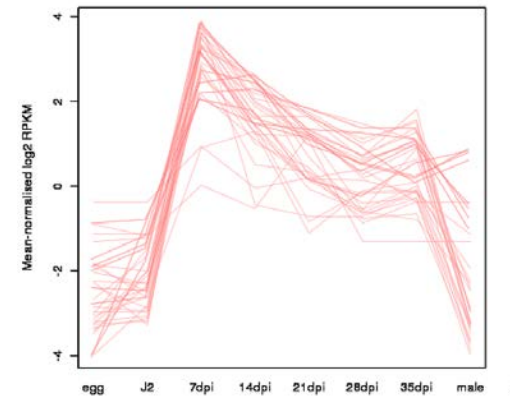
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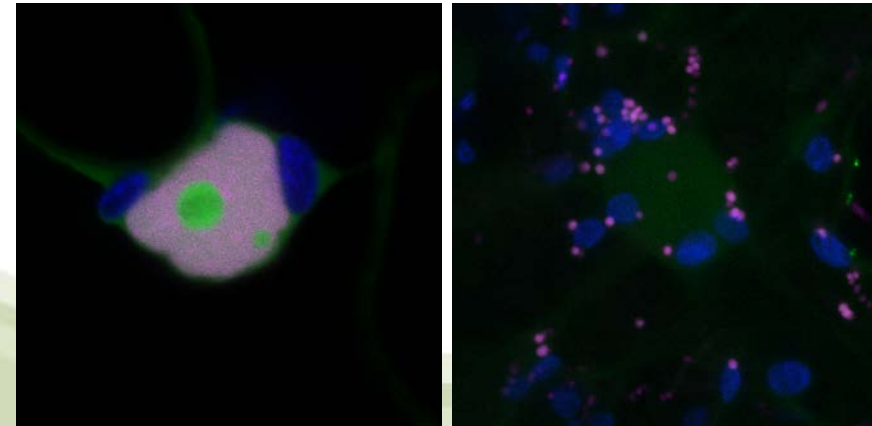
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J2 specific



Parasitic stages



# Some *G. pallida* effectors suppress host defences

## *e.g* SPRYSEC effector SPRY12

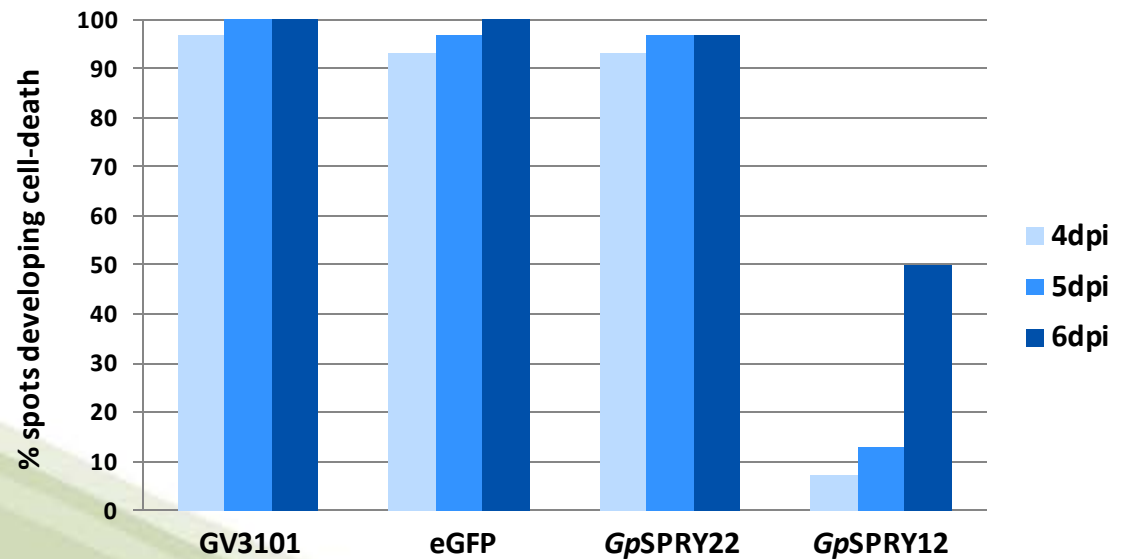
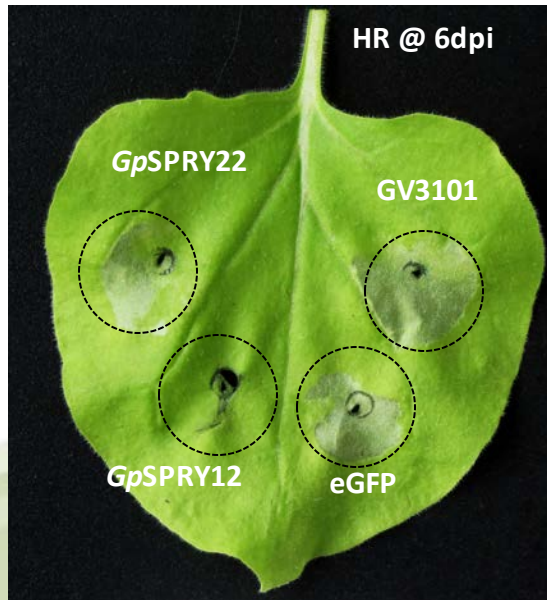
- Expressed in dorsal gland cell
- Suppresses cell death induced by presence of Gpa2/AvrGpa2



# Some *G. pallida* effectors suppress host defences

## e.g SPRYSEC effector SPRY12

- Expressed in dorsal gland cell
- Suppresses cell death induced by presence of Gpa2/AvrGpa2



An ability shared with other plant pathogens

# Some *G. pallida* effectors may manipulate the host as a food source

## SPRYSEC effector SPRY17 interacts with potato CCD4

- Strong interaction in yeast
- BiFC shows that the interaction occurs in the cytoplasm of plant cells

## ■ Reporter *LacZ*

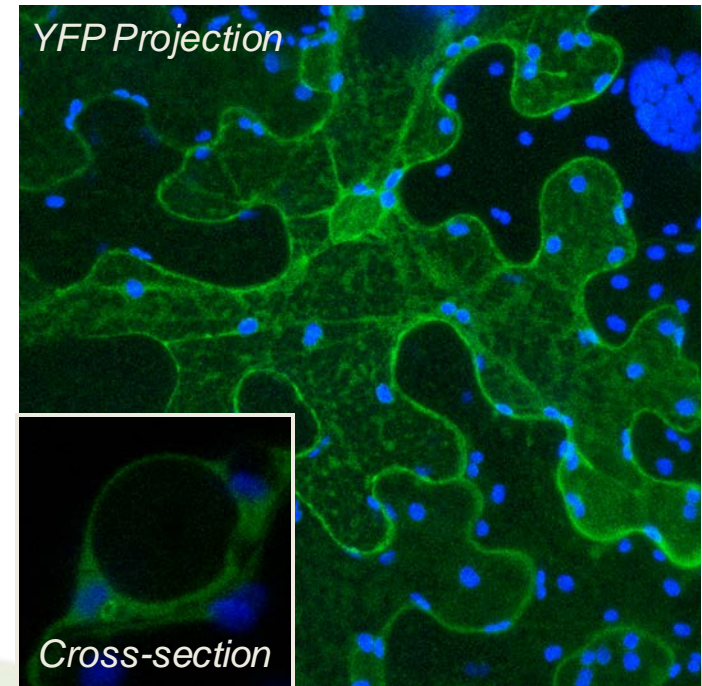
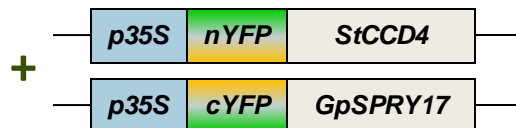
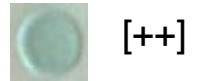
GpSPRY17 + Empty prey vector



StCCD4 + Empty bait vector



GpSPRY17 + StCCD4



BiFC signal

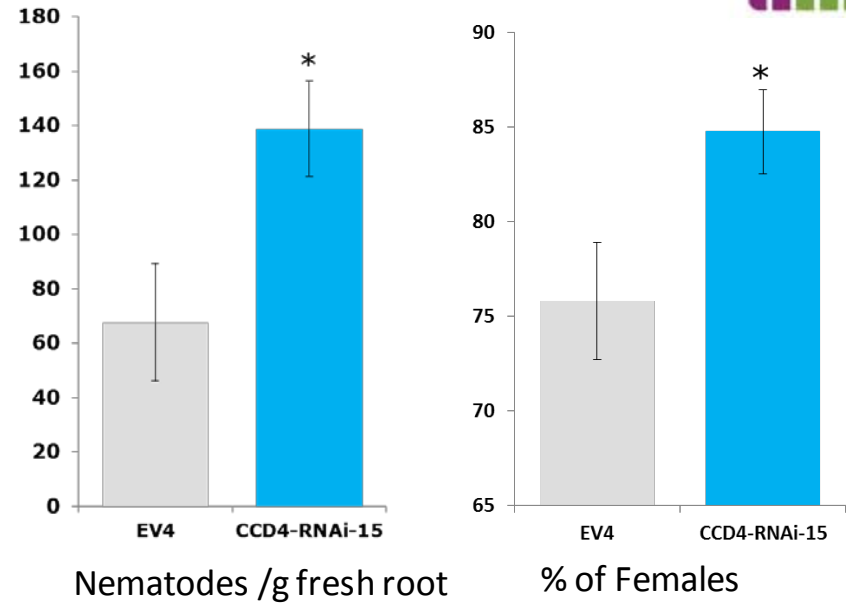


# Silencing CCD4 makes plants more susceptible to *G. pallida*



## CCD4 is a carotenoid cleavage dioxygenase

- Silencing of CCD 4 leads to increased carotenoid levels in plants
- May be a requirement in the *G. pallida* diet

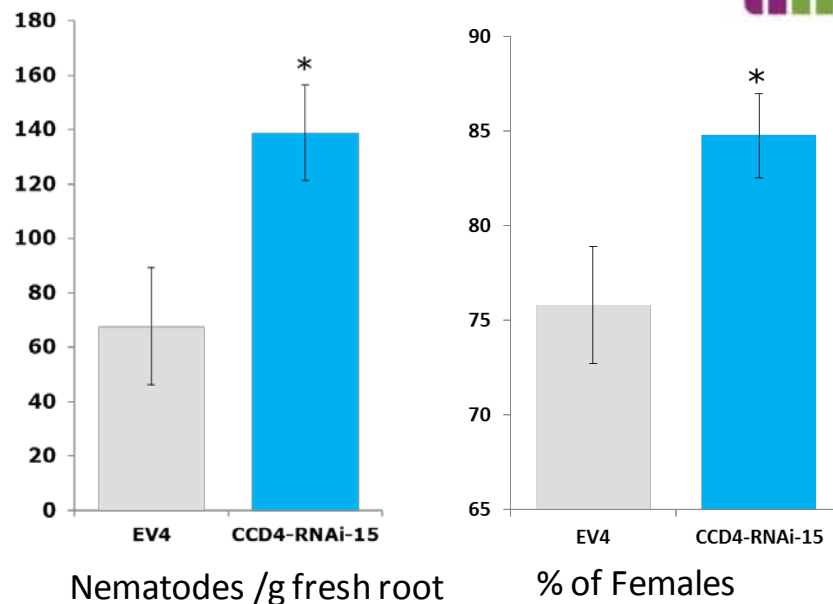


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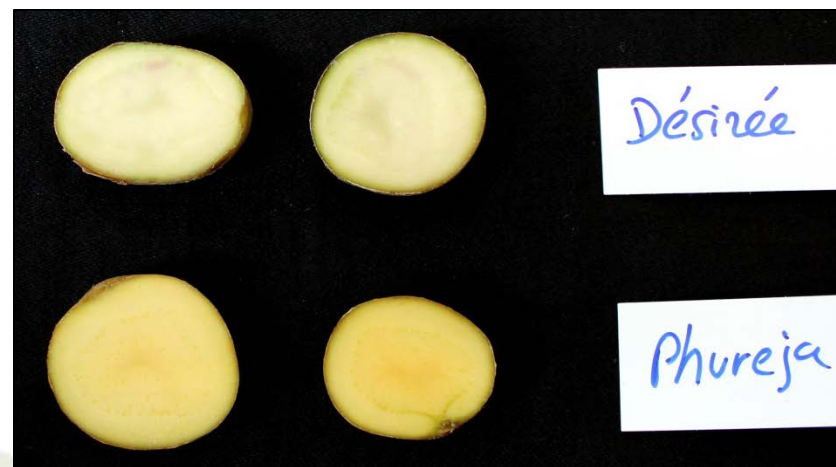
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## A similar pattern is seen in *Solanum phureja*

- Naturally low levels of CCD4
- High levels of carotenoids
- **Highly susceptible to PCN**

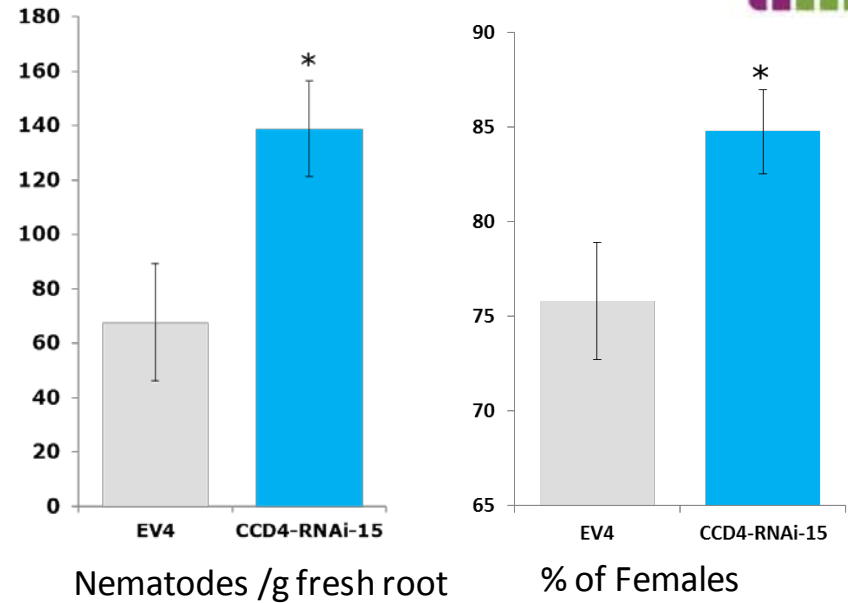


# Silencing CCD4 makes plants more susceptible to *G. pallida*



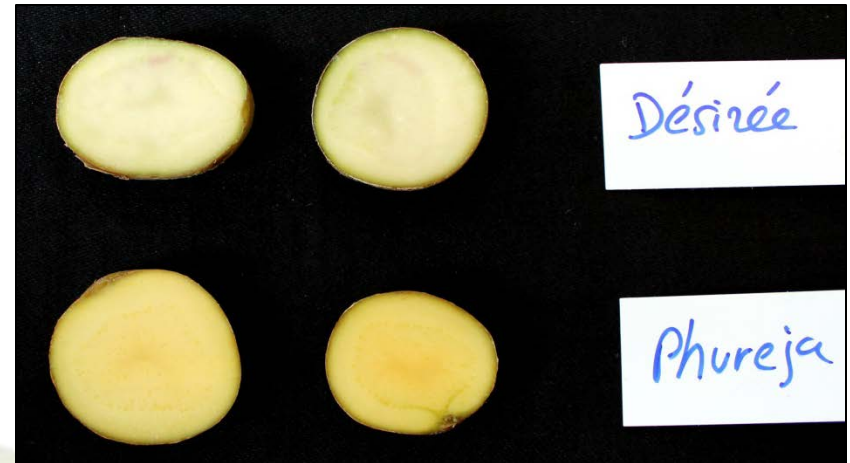
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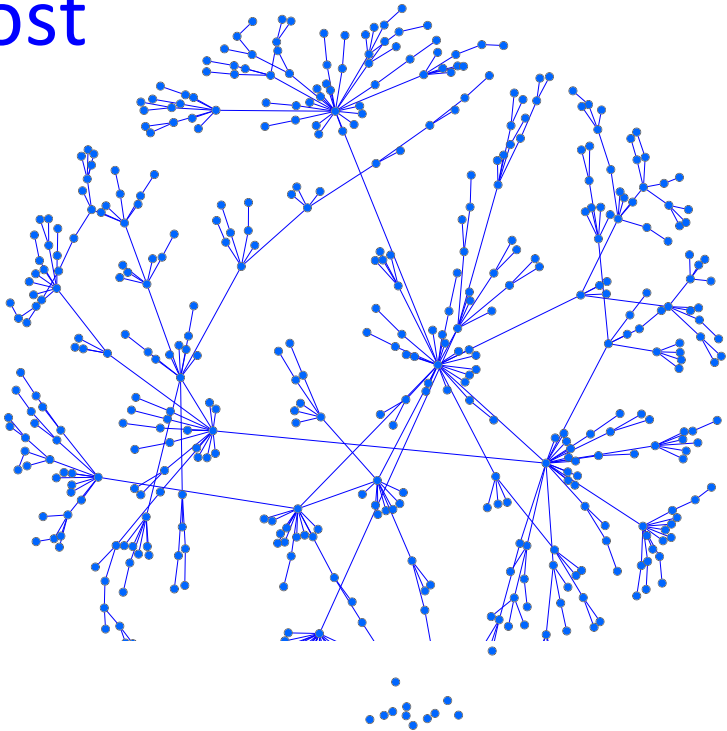
*G. pallida* may inactivate CCD4 in order to improve the nutritional quality of its host

# Common host targets of effectors

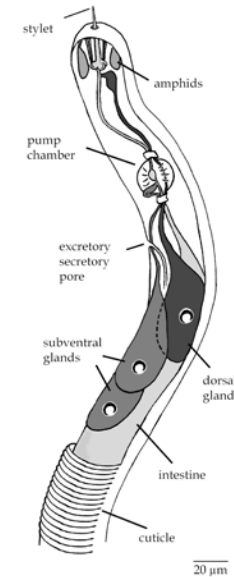
Large scale interaction studies show that some plant proteins interact with many other proteins

- “Hubs”
- Key roles in controlling biochemical networks
- These hubs are frequently targeted by pathogens – control targets

Host



Pathogen

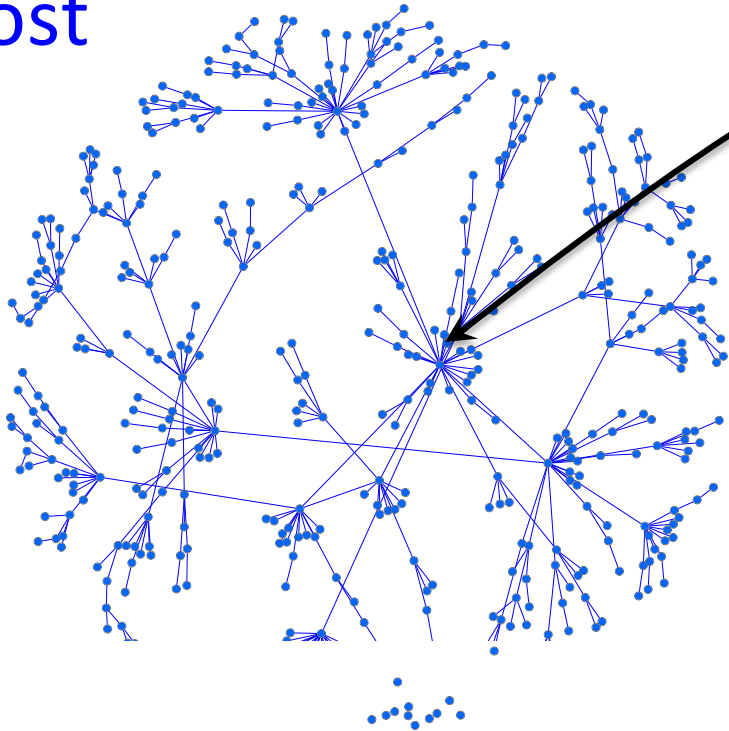


# Common host targets of effectors

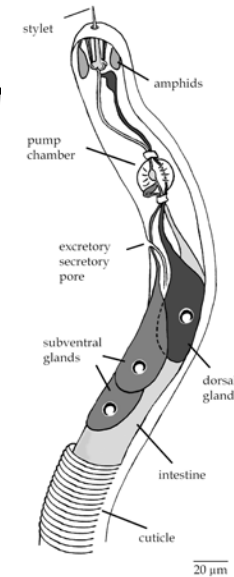
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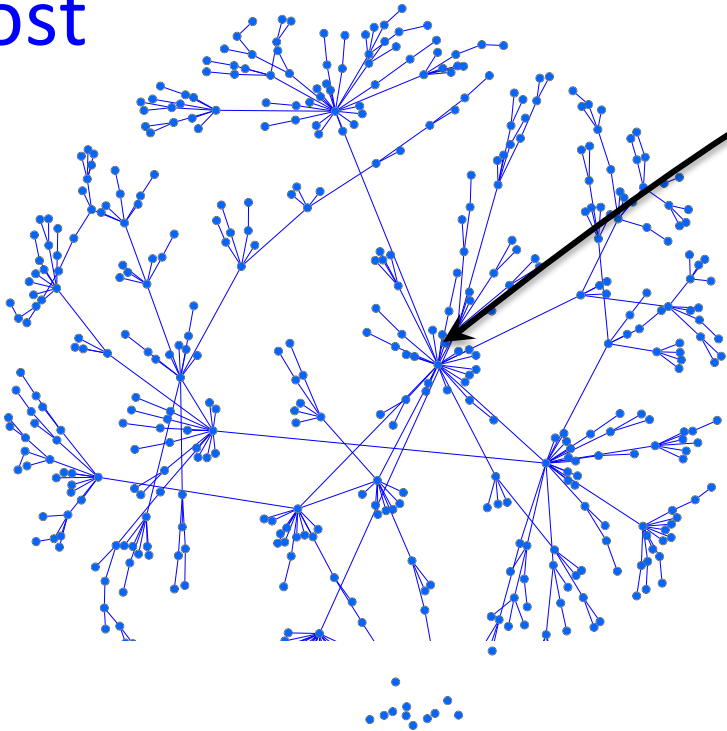


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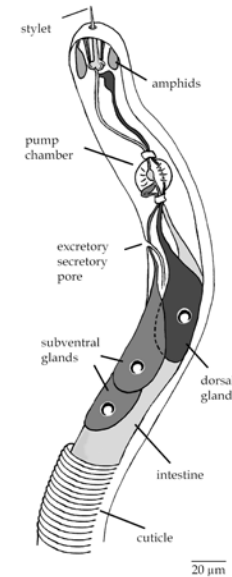
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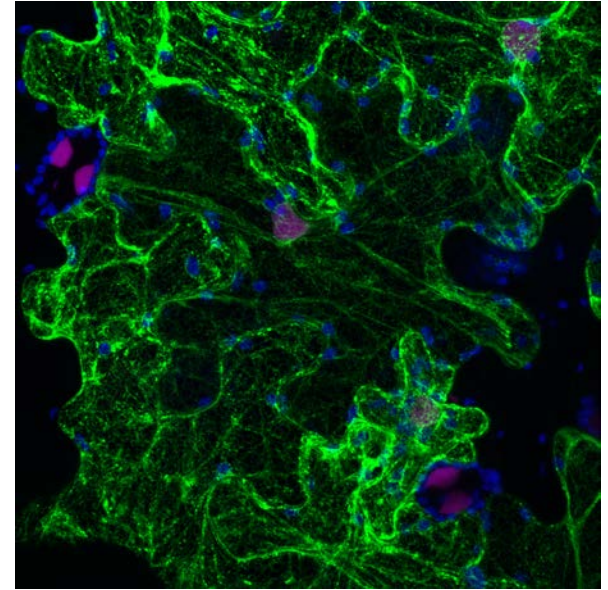
Do *G. pallida* effectors target similar proteins to other pathogens?

# *G. pallida* effector G16H02



## Localises to endoplasmic reticulum in host

- Similar localisation seen for one *Phytophthora infestans* effector
- Interacts with a NAC transcription factor



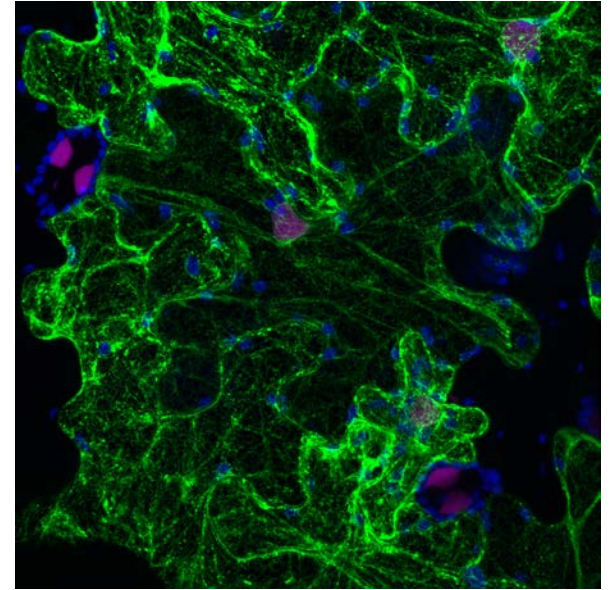
# *G. pallida* effector G16H02



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***G. pallida* G16H02 effector also interacts with a NAC transcription factor**





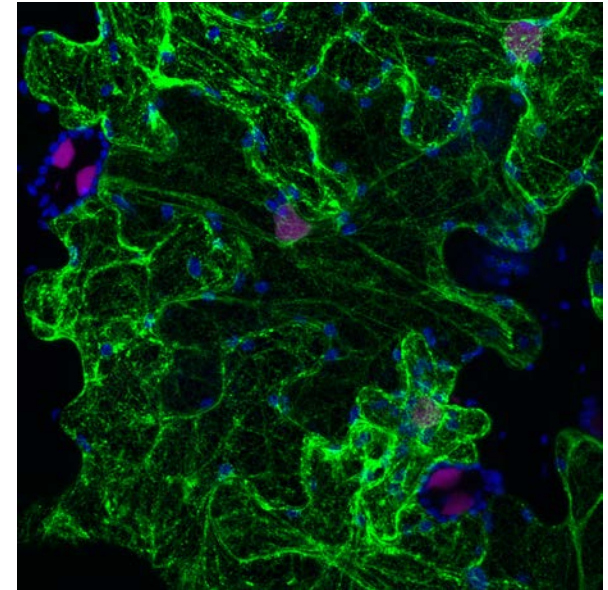
# *G. pallida* effector G16H02



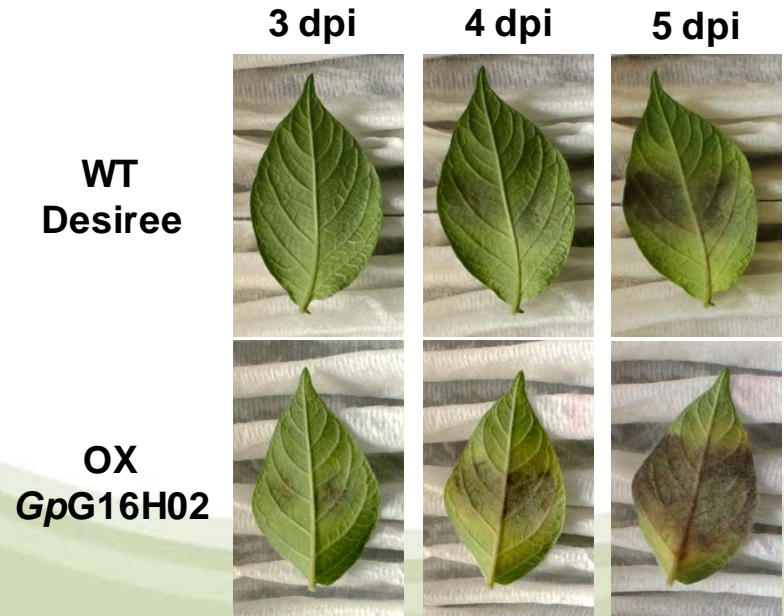
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## *G. pallida* G16H02 effector also interacts with a NAC transcription factor



## Over expression allows increased infection by *P. infestans*

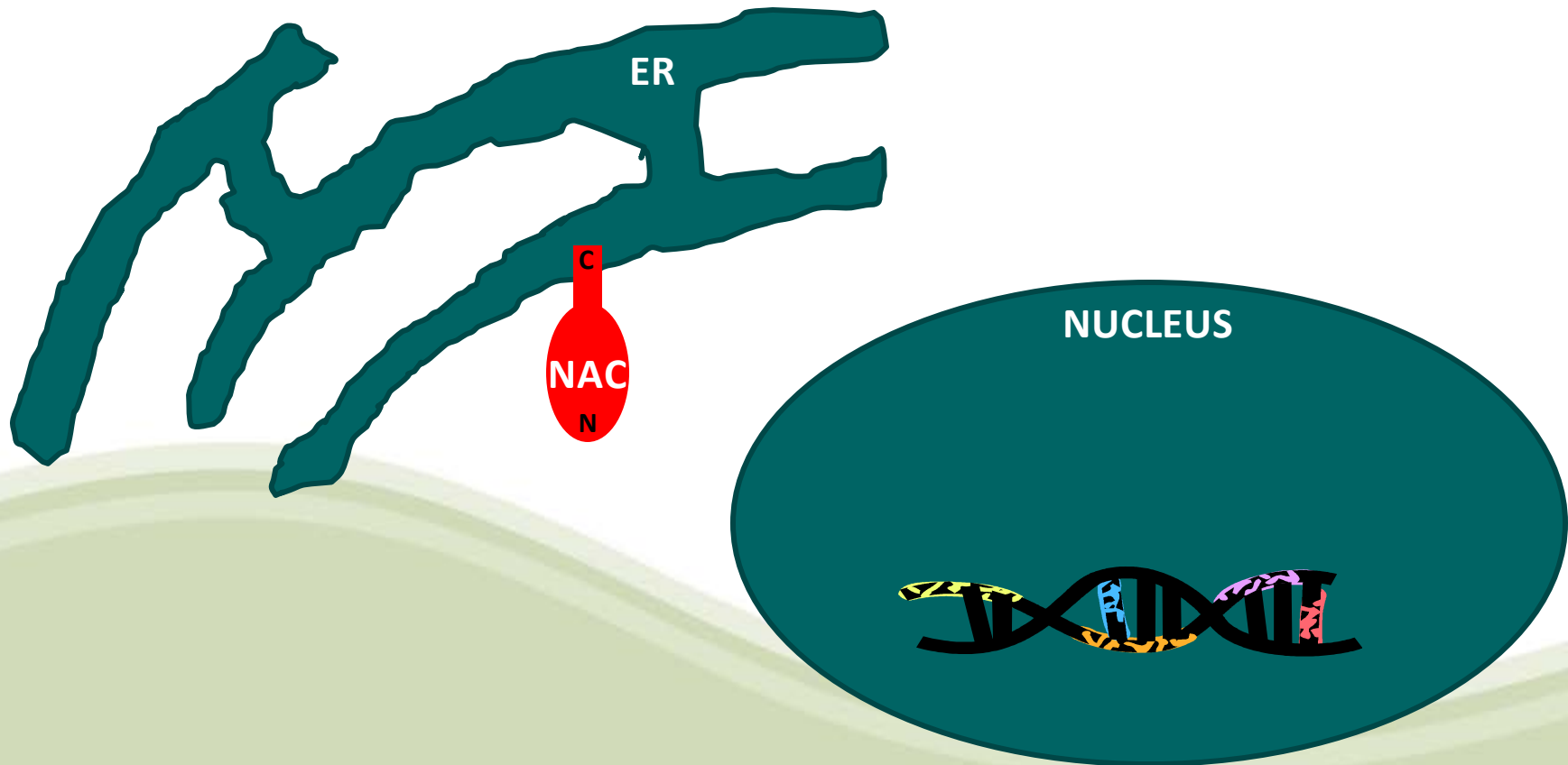


# What are NAC Transcription Factors?

One of largest families of plant-specific TFs

Some are membrane bound TFs (MTFs)

MTFs associated with environmental stress responses

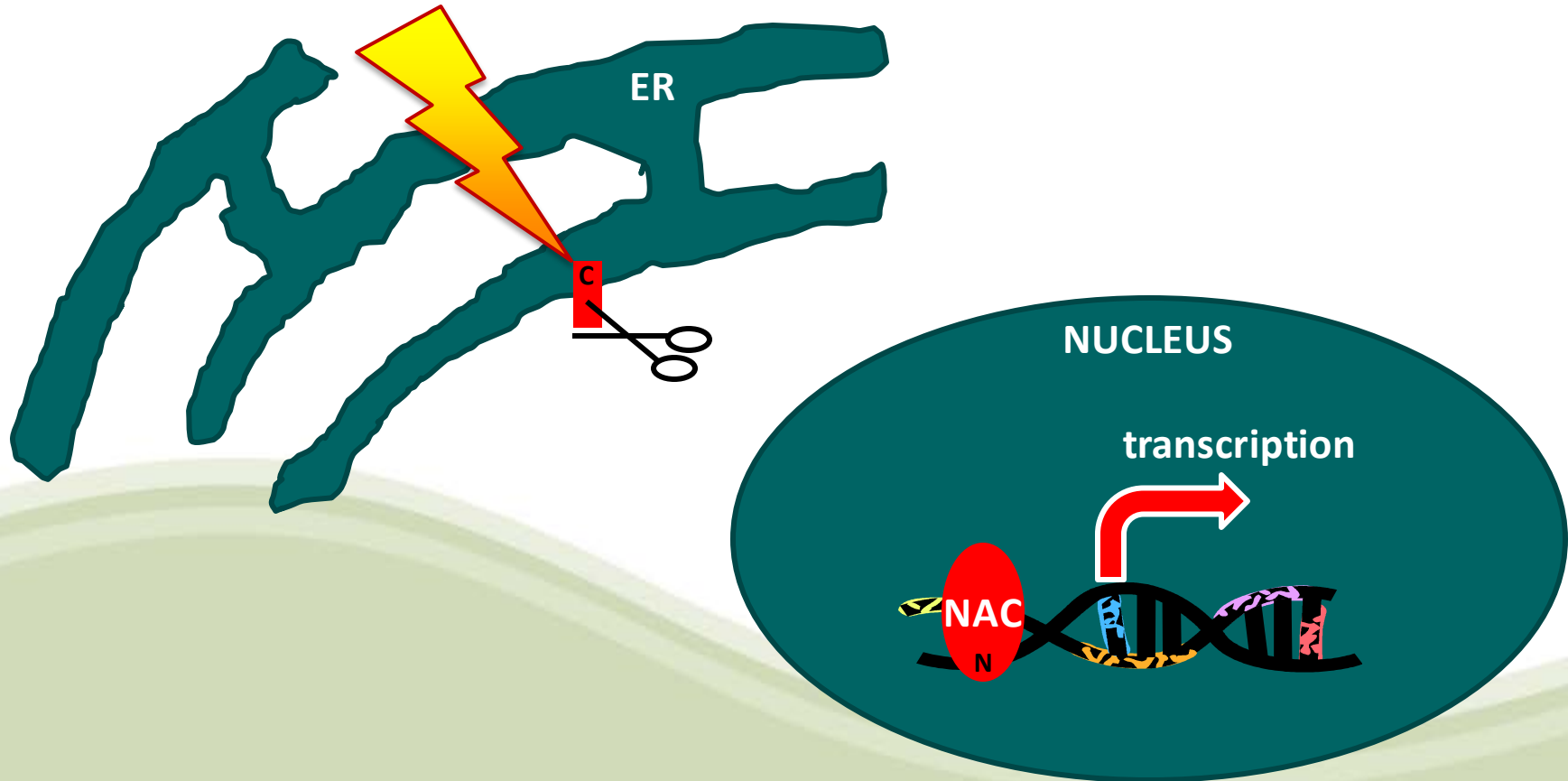


# What are NAC Transcription Factors?

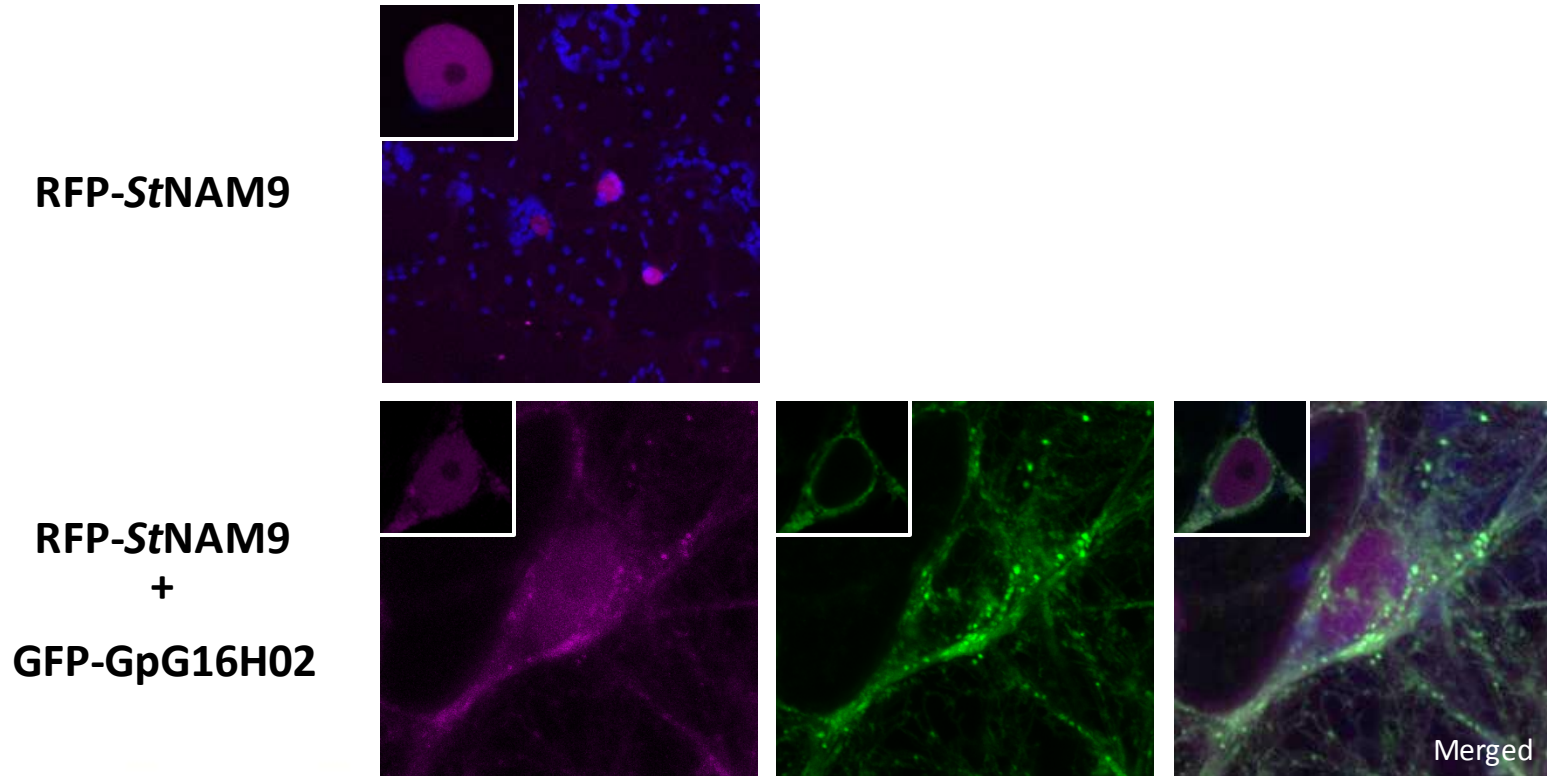
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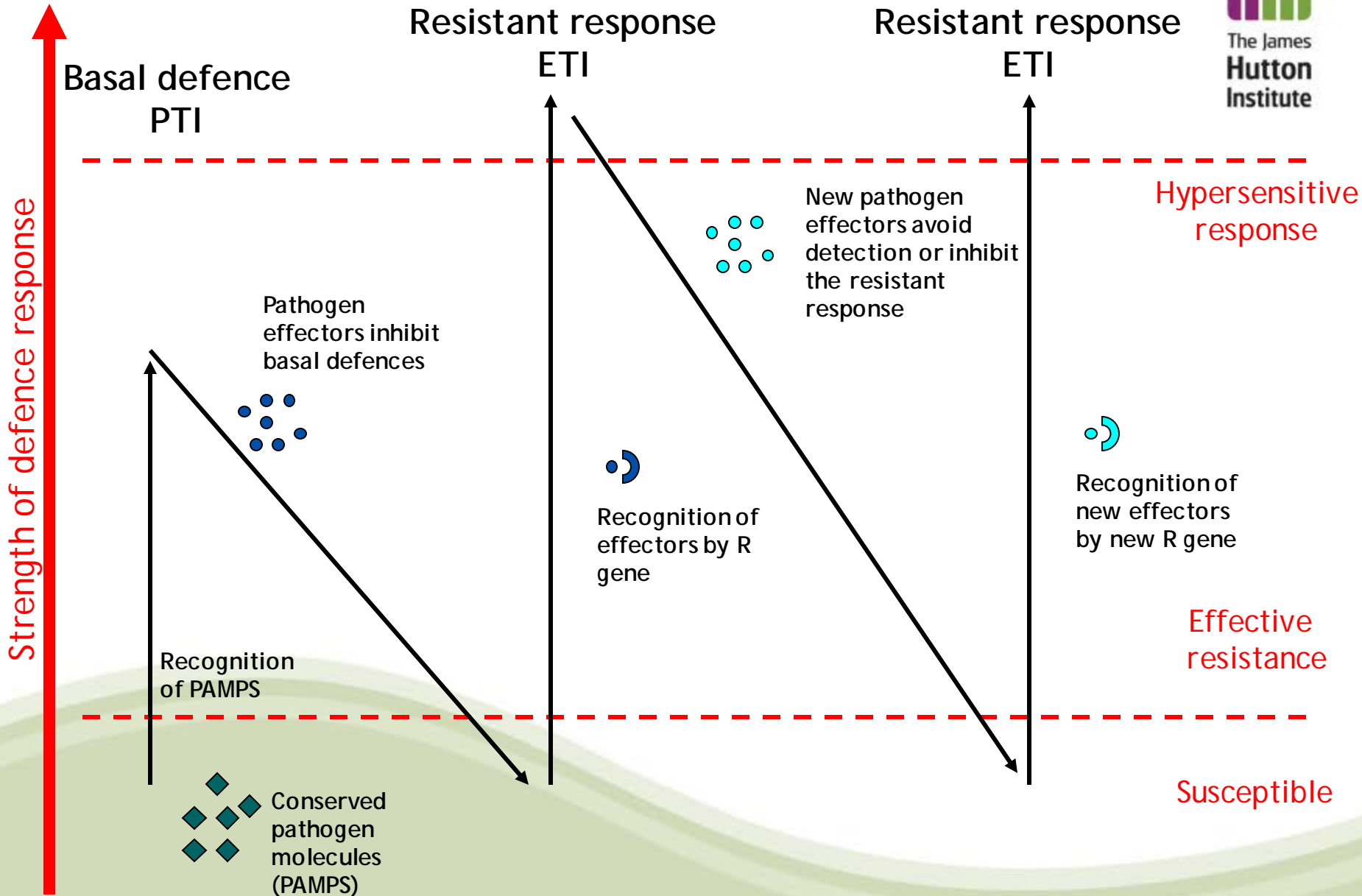
# The *G. pallida* G16H02 effector causes retention of the NAC TF in the ER



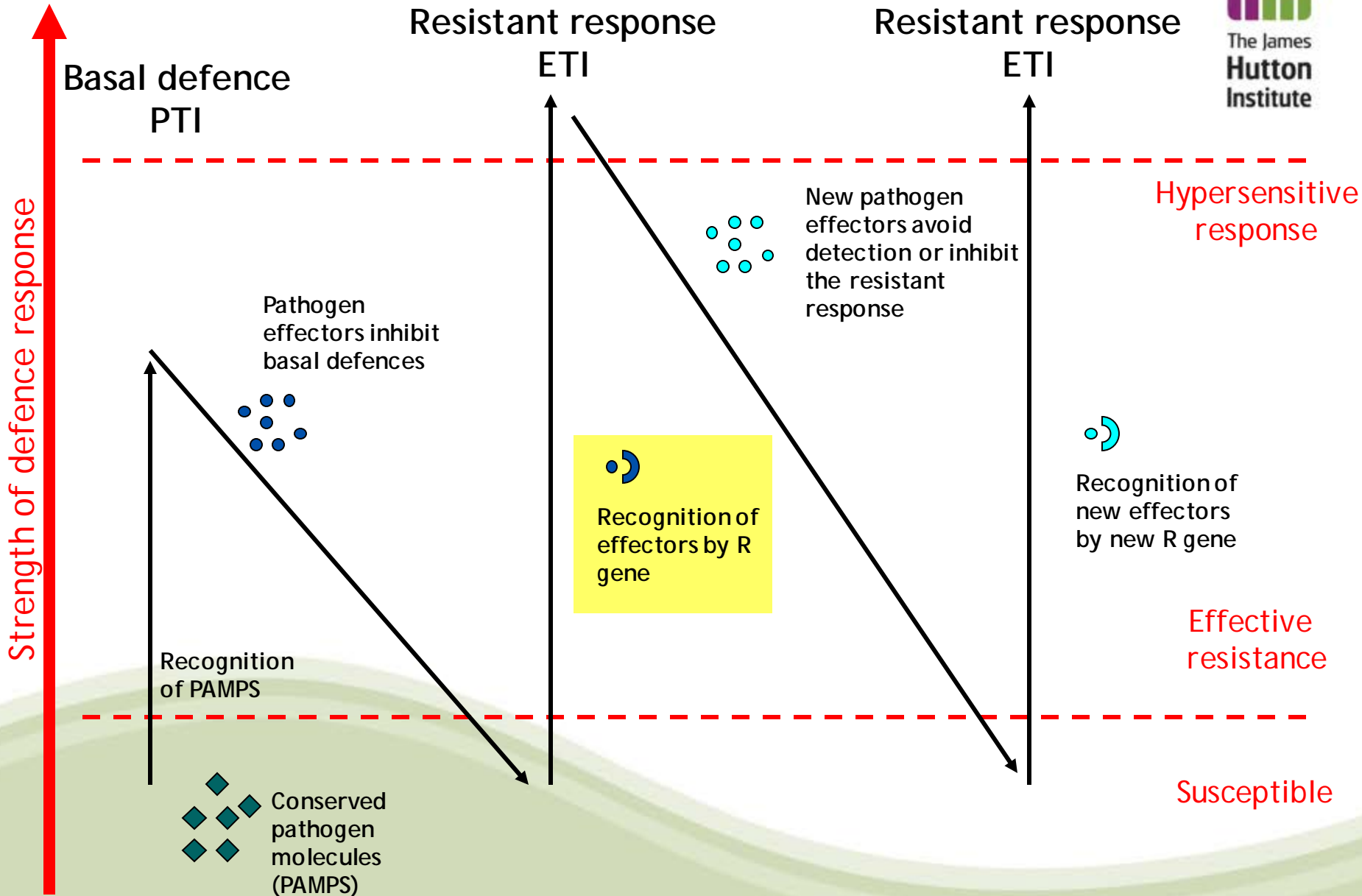
Similar mode of action to the *P. infestans* effector

- Conserved targets of effectors from diverse pathogens

# Identification of effectors: what's the point?



# Identification of effectors: what's the point?



# Recognised effectors can be used to screen for resistance



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## Some effectors are recognised by resistance genes

- Avirulence genes
- SPRYSEC protein – RBP1 = AvrGpa2

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## Some effectors are recognised by resistance genes

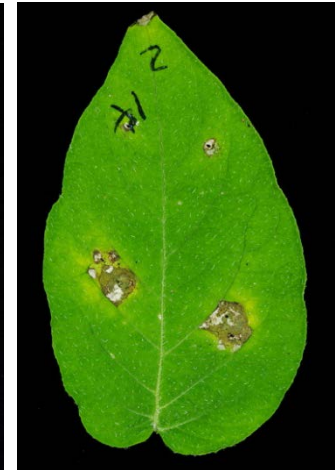
- Avirulence genes
- SPRYSEC protein – RBP1 = AvrGpa2

## Avr genes can be used to screen for resistance in breeding lines

- Assays on leaves
- Transient expression of RBP1



Desiree (Gpa2-)



Cara (Gpa2+)



# Identifying Avr genes for new R sources



## Additional *G. pallida* sequence resources generated

“Population”	Potential uses	gDNA	RNAseq
Bedale (UK <i>G. pallida</i> )	Different virulence compared to sequenced strain	😊	😊
Newton (UK <i>G. pallida</i> )	Starting population for selection	😊	😊
Selected lines	Selected for virulence against <i>verneii</i> and <i>andigena</i>	😊	😊
Luffness	Virulent against <i>verneii</i>	😊	😊
Pa1	Avirulent against H2	😊	😊
P4A (S. American <i>G. pallida</i> )		😊	😊
P5A (S. American <i>G. pallida</i> )	Virulent against <i>verneii</i> and <i>andigena</i> sources	😊	😊

**Analyse variation in effector sequences associated with differences in virulence**

**New tools for resistance breeding**

**Resistance that recognises conserved and/or essential effectors is likely to be durable**

# Summary

Effectors from *G. pallida* and their host targets have been identified

Effectors localise to a range of host structures

*G. pallida* effectors suppress host defence responses

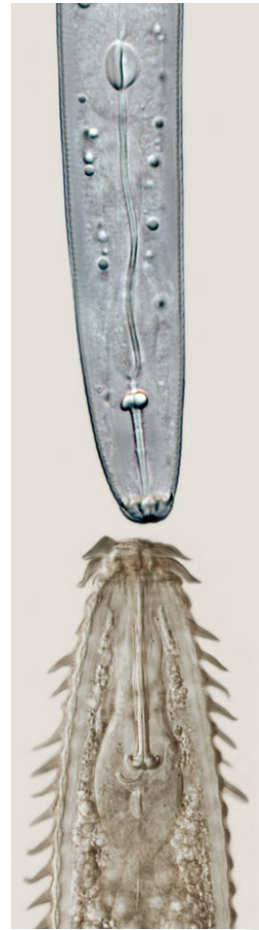
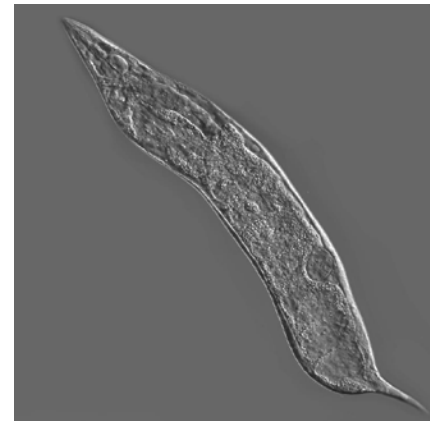
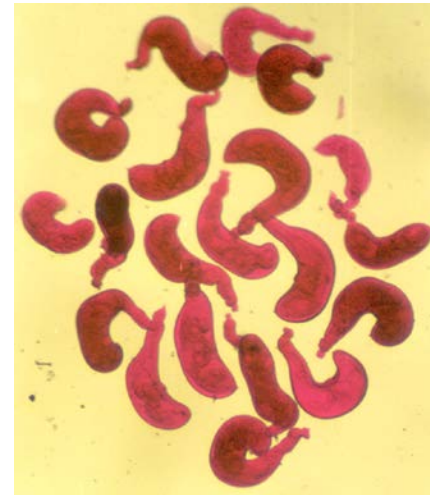
A *G. pallida* effector may manipulate host biochemistry to improve the nutritional qualities of its host

Effectors from diverse pathogens may have similar host targets

Effectors can be used as tools for identifying resistance



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# Acknowledgements

## Nematology team

Vivian Blok

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Ulrike Garnter

Xuan Wang

Jamie Orr

Pete Thorpe

Sophie Mantelin

Aamir Khan

Anne Holt

Kath Wright

Shona Strachan

Dom Laetsch

## Dundee Effector Consortium

### Collaborators

Leeds University (P. Urwin, Catherine Lilley)

Ghent University (Godelieve Gheysen)

University of Edinburgh (M. Blaxter)

